

Structure Human chromodomain Y like 2, chromodomain, in complex with peptido-mimetic UNC3866

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Gro CDYL2 was expressed in E.coli BL21 (DE3) codon plus RIL in Terrific Broth (TB) in the presence of
with 50 µg/mL of kanamycin. Cell were grown at 37 °C to an OD600 of 1.5 and induced by isopropyl-1-
meth thio-D-galactopyranoside (IPTG), final concentration 0.2 mM, and incubated overnight at 16 °C. Cell
od pellets collected by centrifugation and frozen at -80 °C.

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Wash buffer: 20 mM Tris pH 7.5, 400 mM NaCl, **5% glycerol** and 25 mM imidazole;

Elution buffer: 20 mM Tris pH 7.5, 400 mM NaCl, **5% glycerol** and 300 mM imidazole;

Gel filtration buffer: 20 mM Tris-HCl pH 7.5, 100 mM NaCl and 1 mM DTT.

Purification procedure The fusion proteins were purified by Ni-NTA agarose column. The His tag was cleaved by His-tagged TEV protease (purified in-house) with an approximate molar ratio of 1 : 20 in the dialysis buffer (20 mM Tris-HCl, pH 7.5, 150 mM NaCl and 5 mM beta-mercaptoethanol) at 4 °C overnight. The protein was diluted and applied onto HiTrap Q chromatography column (GE Healthcare) equilibrated with 20 mM Tris-HCl pH 7.5, 25mM NaCl and 1mM DTT. The proteins were eluted with a linear gradient of 0-50% elution buffer (20 mM Tris-HCl pH 7.5, 1M NaCl and 1 mM DTT). The proteins were further purified by gel filtration Superdex 200 10/300 (GE Healthcare). The gel filtration buffer contains 20 mM Tris-HCl pH 7.5, 150 mM NaCl and 1 mM DTT.

Protein stock concentration The purified protein was concentrated to 18 mg mL⁻¹.

Crystallization Protein was incubated with UNC3866 at a molar ratio of 1:1.5 for 1 h on ice before setting up the crystallization trial. The crystals were obtained in 25% P3350, 0.2M ammonium acetate, 0.1M HEPES, pH 7.5. The crystals were cryo-protected in the reservoir solution supplemented with 20% (v/v) glycerol and flash-frozen in liquid nitrogen.