

Molecular Biology

Entry Clone Source: Complex

SGC Construct ID: XX01SUCLG1A-c001

Protein Region: None-None

Vector: pNic-Bsa4

Tag: N-6HIS;N-TEV

Host: BL21(DE3)-R3-pRARE2

Note this plasmid encodes two genes (below) interspersed by a ribosomal binding site.

Sequence (with tag(s)):

SUCLG1A:

MHHHHHSSGVDLGTENLYFQSMSYTASRQHLYVDKNTKIICQGFTGKQGTFFHSQQALE
YGTKLVGGTTPGKGGQTHLGLPVFNTVKEAKEQTGATASVIYVPPFAAAINEAIEAEIP
LVVCITEGIPQQDMVRVKHKLLRQEKTRLIGPNCPGVINPGECKIGIMPGHIHKKGRIGIVS
RSGTLTYEAVHQTQVGLGQSLCVGIGGDPFNGTDFIDCLEIFLNDSETEGIIIGEIGGNA
EENAAEFLKQHNSGPNKPVVSFIAGLTAPPGRRMGHAGAIAGGKGGAKEKISALQSAG
VVVSMSPAQLGTTIYKEFEKRKML

SUCLA2A:

MNLSLHEYMSMELLQEAGVSVPKGYVAKSPDEAYAIKKLGSKDVVIKAQVLAGGRGK
GTFESGLKGGVKIVFSPEEAKAVSSQMIGKKLFTKQTGEKGRICNQVLVCERKYPRREYY
FAITMERSFQGPVLIGSSHGGVNIEDVAAESPEAIKEPIDIEEGIKKEQALQLAQKMGFPPN
IVESAAENMVKLYSLFLKYDATMIEINPMVEDSDGAVLCMDAKINFDSNSAYRQKKIFDL
QDWTQEDERDKDAAKANLNYIGLDGNIGCLVNGAGLAMATMDIIKLHGGTPANFLDVG
GGATVHQVTEAFKLITSDKKVLAILVNIFFGIMRCDVIAQGIVMAVKDLEIKIPVVVRLQG
TRVDDAKALIADSGLKILACDDLDEAARMVVKLSEIVTLAKQAHVDVKFQLPIWQ

Protein Expression

Medium: Terrific broth

Antibiotics: Kanamycin, Chloramphenicol

Procedure: Protein was expressed in BL21(DE3)-R3-pRARE2 for 16 hr at 4°C after induction with 200 µM IPTG at an OD of 0.6 measured at 600nm.

Protein Purification

Procedure: Extraction Buffer: 500 mM NaCl, 50 mM HEPES pH 7.5, 20 mM imidazole, 0.5 mM TCEP, 5% glycerol, 1:1000 of Merck Protease Cocktail II.

Binding Buffer: 500 mM NaCl, 50 mM HEPES pH 7.5, 20 mM imidazole, 0.5 mM TCEP, 5% glycerol.

Washing Buffer: 500 mM NaCl, 50 mM HEPES pH 7.5, 40 mM imidazole, 0.5 mM TCEP, 5% glycerol.

Elution Buffer: 500 mM NaCl, 50 mM HEPES pH 7.5, 250 mM imidazole, 0.5 mM TCEP, 5% glycerol.

GF buffer: 500 mM NaCl, 50 mM HEPES pH 7.5, 0.5 mM TCEP, 5% glycerol

A 6L cell pellet was re-suspended in 120ml extraction buffer. Cells were lysed via sonication. The cell debris was removed via centrifugation using a JA 17.00 rotor at 35K for 1hr at 4°C. The supernatant was incubated with 2.5 ml Ni-NTA for 1 hr. Ni-NTA resin bound to the target

proteins was washed two times with 50 ml of binding buffer followed by two times with 20 ml of wash buffer and eluted in 5 ml fractions with elution buffer. Elution's containing protein were loaded onto a S200 gel filtration column and purified via size exclusion. The protein was subsequently incubated with TEV protease at a ratio of 20:1 mg/ml (protein:TEV) overnight at 4°C. Cleaved protein was passed back over 1 ml of Ni-NTA and the flow through containing protein was concentrated to 7 mg/ml.

Columns: Column 1: 2 ml Ni; Column 2: S200 ; Column 3: Ni rebind

Concentration: 7.0 mg/ml

Structure Determination

Crystallization: Crystals were grown by vapour diffusion method. sitting drops containing 100 nL protein (6.2 mg/mL) and 50 nL well solution containing 2.1M DL- malic acid were equilibrated at 20°C.

Data Collection: *Beamline:* Dmnd I04; *Resolution:* 2.59 Å

Data Processing: Data was processed using Xia2 auto processing pipeline. The protein crystallised in the trigonal [P3₁21](#) space group and contained 1 copy of each subunit in the asymmetric unit. The structure was solved via molecular replacement with 1EUC. The final model was produced by iterative cycles of restrained refinement and model building using REFMAC5 and COOT.