

Structure	MBD1
PDB Code	6D1T
Entry clone accession	AF078830
Entry clone source	AU94-F10
Tag	N-terminal hexahistidine tag with integrated TEV protease cleavage site: mhyyyyhhsgrenlyfqq. mhyyyyhhsgrenlyfqq
Construct sequence	MAEDWLDCPALGPFWKRREVFRKSGATGRSDTYYQSPTGDRIRSKVELTRYLGPACDLTLFDFKQGII
Vector	pET28-MHL
Expression host	<i>E. coli</i> BL21(DE3)-V2R-pRARE2
Growth medium	TB A fresh transformation was used to inoculate 50 mL LB media containing 50 µg/mL kanamycin and 30 µg/mL Chloramphenicol. The culture was grown overnight at 37°C with shaking. The next day this starter culture was used to inoculate 2 L of TB growth medium. The culture was grown in LEX at 37 °C to OD ₆₀₀ of 1.0.
Growth method	IPTG-based induction was carried out according to the manufacturer's protocol. The temperature was reduced to 16 °C and the culture was incubated for a further 18 hours before harvesting the cells.
Extraction buffers	Lysis buffer: 20 mM, Tris pH 7.5, 500 mM NaCl, 5% Glycerol, 0.1% NP40, 1 mM PMSF Cells were harvested by centrifugation and pellets were stored in -80 °C.
Extraction procedure	Prior to purification, the cell pellet was resuspended in lysis buffer. Cells were disrupted by sonication (10 minutes) and samples were centrifuged for 60 min at 70000 g.
Purification buffers	NiNTA Elution buffer (EB): 20 mM Tris-HCl pH 7.5, 500 mM NaCl, 250 mM Imidazole Gel Filtration buffer: 20 mM Tris pH 7.5, 150 mM NaCl, 1 mM DTT. Column 1: Affinity purification, open Ni-NTA column Procedure: The supernatant was incubated with 6mL of 50% slurry Ni-NTA beads by rocking. After 30 min incubation at 4°C, the beads were washed with 50 mL of lysis buffer. The protein was eluted using ~20 mL EB.
Purification procedure	The Column 2: Gel Filtration (Superdex S75 16/60 Hi-Load, GE Healthcare). Then the fractions containing protein were identified on a SDS-PAGE gel. 10 mg/ml. 30% PEG-550-MME, 0.1 M magnesium chloride, 0.1 M HEPES
Protein stock concentration	
Crystallization	
Data collection	