

Stru GID4 fragment in complex with a peptide
cture

PDB

Cod 6CDG, 6CDC, 6CD9, 6CD8, 6CCU, 6CCT
e

Entr

y

clon

e BC041829.1

acce

ssio

n

Entr

y

clon

e MGC:43491 IMAGE:5268071

e

sour

ce

SGC

clon

e JMC130-A04

acce

ssio

n

Tag N-terminal tag: MHHHHHHSSGRENLNFQG

MHHHHHHSSGRENLNFQG

Con

struc SGSKFRGHQKSKGNSYDVEVVLQHVDTGNSYLCGYLKIKGLTEEYPTLTTFFEGEIISKKHPFL
t TRKWDADEDVDRKHWGK

sequ

ence FLAFYQYAKSFNSDDFDYEEELKNGDYVFMWRKEQFLVPDHTIKDISGASFAGFYYICFQKSAA
SIEGYYYHRSSEWYQSLNLTHV

Vect pET28-MHL

or

Expr

essio BL21 (DE3) Codon plus RIL (Stratagene)

n

host

Gro GID4 was expressed in E.coli BL21 (DE3) codon plus RIL in Terrific Broth (TB) in the presence of 50 wth μ g/mL of kanamycin. Cell were grown at 37 °C to an OD600 of 1.5 and induced by isopropyl-1-thio-meth D-galactopyranoside (IPTG), final concentration 0.2 mM, and incubated overnight at 16 °C. Cell od pellets collected by centrifugation and frozen at -80 °C.

Extr

actio Lysis buffer: 20 mM Tris-HCl pH 7.5, 400 mM NaCl, 5% glycerol and 2 mM beta-n buff mercaptoethanol

ers

Extr

actio Frozen cell pellet was thawed and suspended in lysis buffer. The cells were lysed by sonication n (Virtis408912, Virsonic) on ice: the sonication protocol was 5 sec pulse at half-maximal frequency proc (5.0), 7 second rest, for 10 minutes total sonication time per pellet. The lysate was centrifuged at edur 15000rpm for 1h.

e

Purif Wash buffer: 20 mM Tris pH 7.5, 400 mM NaCl, 5% glycerol and 25 mM imidazole;

icati Elution buffer: 20 mM Tris pH 7.5, 400 mM NaCl, 5% glycerol and 300 mM imidazole;

on Gel filtration buffer: 20 mM Tris-HCl pH 7.5, 100 mM NaCl and 0.5 mM TCEP

buff
ers

Purif The fusion proteins were purified by Ni-NTA agarose column. The His tag was cleaved by His-icati tagged TEV protease (purified in-house) with an approximate molar ratio of 1 : 20 in the dialysis on buffer (20 mM Tris-HCl, pH 7.5, 150 mM NaCl and 5 mM beta-mercaptoethanol) at 4 °C proc overnight. The tag and protease were removed by reloading onto the Ni-NTA. The proteins were edur further purified by Superdex 200 10/300 (GE Healthcare). The gel filtration buffer contains 20 e mM Tris-HCl, pH 7.5, 100 mM NaCl and 0.5 mM TCEP

Prot

ein

stoc

k The purified protein was concentrated to 8 mg mL⁻¹ using 15 mL concentrators with a 3,000 molecular weight cut-off (Amicon Ultra-15, UFC900524, Millipore).

conc

entra

tion

The purified GID4 (aa 124-289) proteins were separately incubated with different peptides at a molar ratio of 1 : 1.5 for 1 h on ice before setting up the crystallization trials. The crystals of GID4-PGLW, GID4-PSRW and GID4-PTLV were grown in the precipitant conditions containing 20-25% (v/v) Crys PEG3350, 2-3% (v/v) Tacsimate, pH 7.0 and 0.1 M HEPES, pH 7.5 (Average pH 7.4). The GID4-talliz PGLWKS was crystallized in 20% (v/v) PEG3350, 0.2 M NaBr. The GID4-PSRV was crystallized in ation 30% (v/v) PEG2000 and 0.1 M KSCN. The GID4-PHRV was crystallized in 20% (v/v) PEG3350 and 0.03 M Citric acid. The crystals were protected in cryoprotectant solution consisting of reservoir solution supplemented with 20% (v/v) glycerol or 20% (v/v) ethylene glycol and flash-frozen in liquid nitrogen before data collection.