

= SGC clone accession =
PBC007-B09

= Tag =
N-terminal His6-tag,

= Construct comments =

= Construct sequence =
MHHHHHHSSGRENLIFYQGCCEHHKAMIAGLALLRNPELLEIPLALLVVGLGGSLPLF
VHDHFPKSCIDAVEIDPSMLE
VATQWFGFSQSDRMKVHIADGLDYIASLAGGGEARPCYDVIMFDVDSKDPTLGMSCPPP
AFVEQSFLQKVKSILTPEGVF
ILNLVCRDLGLKDSVLAGLKAVFPLLYVRRIEDEVNEILFCQLHPEQKLATPELLETAQAL
ERTLRKPGRGWDDTYVLSD
MLKTVKIV

DNA sequence has been verified by sequencing

= Vector =
pFBOH-MHL

= Expression host =
Spodoptera frugiperda

= Growth method =
Shaker

The recombinant donor vector pFBOH-MHL-METTL13 was transformed into DH10Bac E. coli cells (Invitrogen) to generate recombinant viral DNA.

Sf9 cells (Invitrogen) were transfected with Bacmid DNA using jetPRIME® transfection reagent (PolyPlus Transfection), and recombinant baculovirus particles were recovered.

The recombinant virus preparation was sequentially amplified from P1 to P3 viral stocks. Sf9 cells grown in HyQ® SFX Insect Serum Free Medium (Fisher Scientific) to a density of 4×10^6 cells/mL and with viability not less than 97% were infected with 10 mL of P3 viral stock for each 1 L of cell culture.

Cell culture medium was collected after 4 days of incubation on a shaker at 150 RPM and 27°C when culture viability dropped to 75-80%.

= Extraction buffers =
50 mM Hepes pH 7.4, 500 mM NaCl, 5% Glycerol

= Extraction procedure =
2L native cell pellet was resuspended in a total volume of 200 ml extraction buffer and the cells disrupted by sonication for
10 mins at 5" on 10" off duty cycle at 108W output power.

= Purification buffers =
Washing Buffer: 50 mM Hepes pH 7.4, 500 mM NaCl, 50 mM imidazole
Elution Buffer: 50 mM Hepes pH 7.4, 500 mM NaCl, 5% Glycerol, 250 mM imidazole

= Purification procedure =

The hexahistidine-tagged Mettl13 protein was isolated using Cobalt-charged TALON resin (Clontech),

followed by size exclusion chromatography Superdex 200 (26 x 60) column, pre-equilibrated with 20mM HEPES, pH 7.4, 150mM NaCl, 2mM TCEP.

The collected protein fractions belonging to a single peak were concentrated up to 10 mg/mL and added S-Adenosyl-L-homocysteine (SAH) at a 1:10 molar ratio.

= Protein stock concentration =

Concentration used for crystallization : 10.0 mg/mL

= Mass spec =

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= Functional multimerization =

Monomer

= Crystallization =

Crystallization of 5WCJ was performed in 96 well vapor diffusion sitting drop plates by mixing equal volumes of protein and reservoir solution at 20°C.

Initial hits were observed in 20% (w/v) polyethylene glycol 8000, 0.2M Ammonium Sulfate, 0.1M Sodium Cacodylate (pH 5.5).

After several seeding processes, diffracting quality crystals were grown in sitting drop vapor diffusion plates by mixing 2 μ l of Mettl13 with 1 μ l of 20% (w/v) polyethylene glycol 3350, 200mM Ammonium Chloride and 200nM seeds.

A 30% (v/v) Glycerol supplemented reservoir solution was used as cryoprotectant and cryocooled in liquid nitrogen.