

# Molecular Biology

**Entry Clone Accession:** NM\_017415.2

**Entry Clone Source:** MGC

**SGC Construct ID:** KLHL3A-c003

**Protein Region:** S298-L587

**Vector:** pNIC28-Bsa4

**Tag:** N-6HIS;N-TEV

**Host:** BL21(DE3)-R3-pRARE2

## Sequence (with tag(s)):

MHHHHHHSSGVDLG TENLYFQSM SLPKVMIVVGGQAPKAIRSVECYDFEEDRWDQIAE  
LPSRRCRAGVVF MAGHVYAVGGFNGSLRVRTVDVYDGVK DQWTSIASMQERRSTLGAA  
VLNDLLYAVGGFDGSTGLASVEAYS YKTNEWFFVAPMNTRRSSVGVGVVEGKLYAVGG  
YDGASRQCLSTVEQYNPATNEWIYVADMSTRRS GAGVGVLSGQLYATGGHDG PLVRKSV  
EVYDPGTNTWKQVADMNMCR RNAGVCAVNGLLYVVGDDGSCN LASVEYYNPVTDK  
WTL LPTNMSTGRSYAGVAVIHKSL

## Sequence after tag cleavage:

SMSLPKVMIVVGGQAPKAIRSVECYDFEEDRWDQIAELPSRRCRAGVVF MAGHVYAVG  
GFNGSLRVRTVDVYDGVK DQWTSIASMQERRSTLGAAVLNDLLYAVGGFDGSTGLASVE  
AYS YKTNEWFFVAPMNTRRSSVGVGVVEGKLYAVGGYD GASRQCLSTVEQYNPATNEW  
IYVADMSTRRS GAGVGVLSGQLYATGGHDG PLVRKSVEVYDPGTNTWKQVADMNMCR  
RNAGVCAVNGLLYVVGDDGSCN LASVEYYNPVTDKWTLLPTNMSTGRSYAGVAVIHK  
SL

## DNA Sequence:

CATATGCACCATCATCATCATCATTCTTCTGGTGTAGATCTGGGTACCGAGAACCTGTA  
CTTCCAATCCATGAGCCTTCCCAAGGTCATGATTGTGGTTGGCGGCCAGGCACCCAAG  
GCAATCCGCAGTGTGGAGTGCTATGATTTTCGAGGAGGACCGGTGGGATCAGATTGCTG  
AGCTTCCTTCCAGAAGATGCAGAGCAGGTGTGGTGTTTCATGGCTGGCCACGTGTATGC  
CGTGGGAGGGTTTAATGGCTCACTGCGGGTGCGGACAGTGGATGTGTATGACGGCGT  
GAAGGACCAGTGGACGTCCATTGCCAGCATGCAGGAGCGCCGGAGCACACTGGGCG  
CAGCGGTGCTCAATGACTTGCTCTACGCAGTGGGAGGCTTTGATGGCAGTACTGGCCT  
AGCATCGGTGGAAGCCTACAGCTACAAGACCAACGAGTGGTTCTTTGTGGCCCCGAT  
GAACACGCGGCGGAGCAGTGTGGGTGTGGGCGTTGTGGAGGGGAAGCTATATGCTGT  
TGGGGGTTATGATGGAGCTTCCCGCCAGTGTCTGAGCACTGTGGAGCAGTACAACCC  
AGCGACCAATGAATGGATATACGTGGCGGACATGAGCACCCGCCGAGTGGCGCAGG  
GGTTGGAGTGCTTAGCGGACAGCTGTACGCCACAGGTGGGCATGATGGGCCTTTGGT  
GAGGAAGAGCGTTGAGGTTTACGATCCTGGAACAAATACCTGGAAGCAAGTGGCAGA  
CATGAACATGTGCCGCGCAACGCAGGGGTCTGTGCAGTAAATGGGCTCCTGTATGTG  
GTTGGAGGGGATGATGGATCCTGCAACTTGGCTTCGGTGGAGTACTACAATCCTGTCA  
CTGACAAATGGACGCTGCTTCCAACGAACATGAGCACGGGGCGGAGCTATGCAGGTG  
TTGCCGTGATTCAACAAGTCCTTGTGACAGTAAAGGTGGATACGGATCCGAA

# Protein Expression

**Medium:** Lysogeny broth

**Antibiotics:** Kanamycin

**Procedure:** KLHL3A-c003 was transformed into BL21(DE3)-R3-pRARE2. 10mL E.coli overnight culture was inoculated into 2L autoclaved LB medium with 50ug/mL of each

Kanamycin and Chloramphenicol. E.coli cells were grown in 37°C with 160RPM shaking until

the OD<sub>600</sub> reached 0.6. Expression was induced with 0.4mM Isopropyl  $\beta$ -D-1-thiogalactopyranoside and cultured overnight in 18°C with 160RPM shaking

## Protein Purification

**Procedure:** After cell lysis with 15min sonication, His-KLHL3 was purified by IMAC and eluted out by imidazole. Eluted fractions were pooled and then gel filtrated. His tag was cleaved by TEV protease. Untagged KLHL3 was purified by reverse IMAC. Purified KLHL3 was concentrated to 10mg/mL using 10kD MWCO concentrator. KLHL3 final stock was buffered in 50mM HEPES pH7.5, 300mM NaCl and 0.5mM TCEP.

**Columns:** Column 1: 5mL Ni column; Column 2: Gel F 16/60 S75; Column 3: 0.5 mL Ni rebind

**Concentration:** 10 mg/ml

**Mass-spec Verification:** Intact mass confirmed. (expected - 31499.6. observed – 31500.4)

## Structure Determination

**Crystallization:** KLHL3 protein was co-crystallized with 4mM WNK3 degron peptide (ECEETEVDQHV). Crystals were yielded after micro-seeding. Seeds were obtained from 8% PEG4000 -- 0.1M acetate pH 4.5 at 4°C in coarse screen. Those early crystals were transferred into an eppendorf tube containing 50  $\mu$ L reservoir solution and a seed bead (Hampton Research), then vortexed for 2 min. Seed-stocks were diluted in 500 fold when in use. The drops were spiked with 20 nL of diluted seed-stock solution. The best-diffracting crystals of the KLHL3 complex were obtained at 4°C by mixing 75 nL of protein with 75 nL of a reservoir solution containing 6% PEG4K -- 0.1M acetate pH 5.1. Prior to vitrification in liquid nitrogen, crystals were cryoprotected by direct addition of reservoir solution supplemented with 25 % ethylene glycol.

**Data Collection:** *Beamline:* Dmnd I03; *Resolution:* 2.8 Å; *Wavelength:* 0.9763Å;

**Data Processing:** The data for KLHL3-WNK3 crystal were processed in software PHENIX version1.9 (Adams, Afonine et al. 2010). Molecular replacement was performed with PHENIX.Phaser-MR using PDB code 4CH9 chain A (Kelch domain of KLHL3) as the model.