

# PUS7

**PDB:**5KKP

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**BC011396

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**N-terminal: 6XHis-tag with integrated TEV protease site: MHHHHHHSSGRENLYFQ\*G

**Host:**E.coli BL21 (DE3) codon plus RIL (Stratagen).

## Construct

**Prelude:**

**Sequence:**

gESFADMMKHGLTEADVGITKFVSSHQGFSGILKERYSDFFVHEIGKDGRISHLNDLSIPVDEEDPSEDIFTVLTAEEKQRLEELQL  
FKNKETSVAIEVIEDTKEKRTIIHQAIKSLFPGLETKTEDREGKKYIVAYHAAGKKALANPRKHSWPKSRGSYCHFVLYKENKDTMD  
AINVLSKYLRVKPNIFSVMGTGDKRAITVQEIAVLKITAQRLAHLNKLMMNFKLGNFSYQKNPLKLGELQGNHFTVVLNITGTDDQ  
VQQAMNSLKEIGFINYYGMQRFGTTAVPTYQVGRAILQNSWTEVMDLILKPRSGAEKGYLVKCREEWAKTKDPTAALRKLPVKRCVE  
GQLLRGLSKYGMKNIVSAFGIIPRNNRLMYIHSYQSYVWNNMVSKRIEDYGLKPVPGDLVLKGATATYIEEDDVNNYSIHDVVMPLP  
GFDVIYPKHKIQEAYREMLTADNLDIDNMRHKIRDYSLSGAYRKIIIRPQNVSWEVVAYDDPKIPLFNTDVDNLEGKTPPVFASEGK  
YRALKMDFSLPPSTYATMAIREVLKMDTSIKNQTLNNTWLR

**Vector:**pET28-MHL

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**PUS7 protein was expressed in E.coli BL21 (DE3) codon plus RIL in M9 minimum medium in the presence of 50 µg/ml of kanamycin. Cell were grown at 37°C to an OD<sub>600</sub> of 0.8 and induced by isopropyl-1-thio-D-galactopyranoside (IPTG), final concentration 1 mM, in the presence of 50 mg/L of SeMet and incubated overnight at 15°C.

## Purification

**Buffers**

**Procedure**

The crude extract was cleared by centrifugation. The lysate was loaded onto 5 ml HiTrap column (GE Healthcare), charged with Ni<sup>2+</sup>. The column was washed with 10 CV of 20 mM HEPES pH 7.4, containing 500 mM NaCl, 50 mM imidazole, 5% glycerol, and the protein was eluted with elution buffer (20 mM HEPES pH 7.5, 500 mM NaCl, 250 mM imidazole, 5% glycerol). The

protein was loaded on Superdex200 column (26x60) (GE Healthcare), equilibrated with 20 mM PIPES, pH 6.5, 250 mM NaCl. The fractions containing PUS7 were pooled and TEV protease was added to remove His-tag. The protein was further purified to homogeneity by ion-exchange chromatography on Source 30S column (10x10) (GE Healthcare), equilibrated with buffer containing 20 mM PIPES, pH 6.5, and eluted with linear gradient of NaCl up to 500 mM concentration (20 CV).

## **Extraction**

### **Buffers**

#### **Procedure**

Cells were harvested by centrifugation at 7,000 rpm. The cell pellets were frozen in liquid nitrogen and stored at -80°C. For purification, the cell paste was thawed and resuspended in lysis buffer (50 mM HEPES, pH 7.4, 0.5 M NaCl, 5 mM imidazole, 2 mM  $\beta$ -mercaptoethanol, 5% glycerol) with protease inhibitor (1 mM phenylmethyl sulfonyl fluoride, PMSF). The cells were lysed by passing through Microfluidizer (Microfluidics Corp.) at 20,000 psi.

**Concentration:** 24.7 mg/mL - Enzymatic treatment: TEV

#### **Ligand**

**MassSpec:** The expected mass for PUS7 is 64415 Da, measured mass for SeMet protein is 65214 Da.

**Crystallization:** Purified PUS7 protein (10.1 mg/mL) was crystallized using sitting drop vapor diffusion method at 20 °C by mixing 1  $\mu$ L of the protein solution with 1  $\mu$ L of the reservoir solution containing 20% PEG3350, 0.2 M sodium formate, pH 7.0

#### **NMR Spectroscopy:**

#### **Data Collection:**

#### **Data Processing:**