

# SIAH2

**PDB:**5H9M

**Entry Clone Accession:**BC013082

**Entry Clone Source:**AT13-G11

**SGC Clone Accession:**YTC020-H07

**Tag:**N-terminal SUMO tag, removed

**Host:**BL21-CodonPlus-RIL

**Prelude:**SIAH2:A131-T321:SUMO

**Vector:**pET28-MKH8SUMO

## Sequence:

gASAVLFPCKYATTGCSLTLHHTEKPEHEDICEYRPYSCPCPGASCKWQGSLEAVMSHLMHAHKSITTLQGEDIVFLATDINLPGAV  
DWVMMQSCFGHHFMLVLEKQEKYEGHQFFAIVLLIGTRKQAEAFAYRLELNGNRRRLTWEATPRSIHDGVAAAIMNSDCLVFDTAI  
AHLFADNGNLGINVTIST

## Growth

**Procedure:** LEX Bubbling

For protein: The target protein was expressed in E. coli by inoculating 30 mL of overnight culture grown in Luria-Bertani medium into a 1 L of TerrificBroth medium in the presence of 50 ug/mL kanamycin and 34 ug/mL chloramphenicol at 37 degree. When the OD<sub>600</sub> of the culture reached ~1.5, the temperature was lowered to 18 degree and the culture was induced with 0.25 mM IPTG. The cells were allowed to grow overnight before harvested by centrifugation (7,000 rpm Beckman JLA-8.1000 rotor 12min) and flash frozen in liquid nitrogen and stored at -80 degree.

## Purification

**Procedure:** The lysate was centrifuged at 16,000 rpm (25,800xg RCF(average) for 60 minutes and transferred the supernatant to equilibrated 6 mL Ni-NTA resin and incubated for 1 hour at 4 degree, then loaded them to Bio-rad open gravity column. The beads in the open column was then washed with 25 mL lysis buffer followed by with 50 mL washing

buffer. Bound proteins were eluted using 15 mL elution buffer. The N-terminal Sumo-tag was removed by overnight incubation with TEV protease (1:30 w/w) at 4°C during dialysis against the dialysis buffer. Uncut proteins and TEV protease were removed by passing the solution through 3 mL Ni-NTA beads, and the target protein was further purified by anion-exchange chromatography on 5 mL HiTrap S column (GE Healthcare). Then the protein was further purified by gel filtration on a HighLoad 16/60 Superdex 200 column (GE Healthcare) pre-equilibrated with gel filtration buffer. Fractions containing target protein were pooled and concentrated by centrifugal filters (Amicon MWCO 10 kDa). The final yield of the protein was about 1.3 mg per litre bacterial culture and the purity is above 99% judging from SDS-PAGE.

## Extraction

**Procedure:** 4L cell pellet was resuspended in a total volume of 400 mL lysis buffer with 1 mM PMSF/Benzamidine freshly added and the cells disrupted by sonication for 10 mins at 5" on 7" off duty cycle at 120W output power,

**Concentration:** 11.15 mg/mL

## Structure Determination

**MassSpec:** native protein without SUMO tag measured: 21354.7 g/mol, expected 21354.4 g/mol.

**Crystallization:** The crystals were grown at 298K using the sitting drop method by mixing 0.5 µL protein with 0.5 µL well solution consisting of 25% PEG8000, 0.2 M NaCl, 0.1 M Hepes pH 7.5. The crystals were cryoprotected by immersion in well solution containing 15% ethylene glycol firstly, then immersion in Paratone.