

YANK1A (4FR4) Materials & Methods

Entry Clone Source: Synthetic DNA

SGC Construct ID: YANK1A-c051

Vector: pFB-LIC-Bse. Details [[PDF](#)]; Sequence [[FASTA](#)] or [[GenBank](#)].

DNA sequence:

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ATGGGCCACCATCATCATCATCATTC  
TTCTGGTAGATCTGGTACCGAGA  
ACCTGTACTCCAATCCATGCCACCA  
GTGTTGATGAAATGAAGATGTCAA  
CTTGACCACTTGAAATTTGCGAG  
CCATTGGAAAGGCAGTTGGGAAG  
GTCTGCATTGTACAGAAGAATGATA  
CAAGAAGATGTACGCAATGAAGTACA  
TGAATAAACAAAAGTGCCTGGAGCGC  
AATGAAGTGAGAAATGTCTCAAGGA  
ACTCCAGATCATGCAGGGTCTGGAGC  
ACCCTTCCTGGTTAATTGTGGTAT  
TCCTTCCAAGATGAGGAAGACATGTT  
CATGGTGGTGGACCTCCTGCTGGGTG  
GAGACCTGCGTTATCACCTGCAACAG  
AACGTCCACTTCAAGGAAGAACAGT  
GAAGCTCTTCATCTGTGAGCTGGTCA  
TGGCCCTGGACTACCTGCAGAACCCAG  
CGCATCATTACAGGGATATGAAGCC  
TGACAATATTTACTTGACGAACATG  
GGCACGTGCACATCACAGATTCAAC  
ATTGCTGCGATGCTGCCAGGGAGAC  
ACAGATTACCACCATGGCTGGCACCA  
AGCCTTACATGGCACCTGAGATGTT  
AGCTCCAGAAAAGGAGCAGGCTATT  
CTTGCTGTTGACTGGTGGTCCCTGG  
GAGTGACGGCATATGAAGTGTGAGA  
GGCCGGAGACCGTATCATATTGCTC  
CAGTACTTCCAGCAAGGAATTGTAC  
ACACGTTGAGACGACTGTTGTAAC  
TACCCCTCTGCCTGGTCACAGGAAAT  
GGTGTCACTCTTAAAAAGCTACTCG  
AACCTAATCCAGACCAACGATTTCT  
CAGTTATCTGATGTCCAGAACTTCCC  
GTATATGAATGATATAAACTGGGATG  
CAGTTTTCAGAAGAGGCTCATTCCA  
GGTTTCATTCTAAATAAGGCAGGCT  
GAATTGTGATCCTACCTTGAACCTG  
AGGAAATGATTGGAGTCCAAACCT  
CTACATAAGAAAAAAAGCGTCTGGC  
AAAGAAGGAGAAGGATATGAGGAAAT  
GCGATTCTCTCAGACATGTCTTCTT  
CAAGAGCACCTTGACTCTGTCCAGAA
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GGAGTTCATATTTCAACAGAGAAA
AAGTAAACAGGGACTTAACAAAAGA
CAACCAAATCTAGCCTTGGAACAAAC
CAAAGACCCACAAGGTGAGGATTGA

Final protein sequence (Tag sequence in lowercase):

mghhhhhhsgvdlgtenlyfq^smp
PVFDENEDVNFDHFEILRAIGKGSFG
KVCIVQKNDTKMYAMKYMNKQKCVE
RNEVRNVFKELQIMQGLEHPFLVNLW
YSFQDEEDMFMVVVDLLGGDLRYHLQ
QNVHFKEETVKLFICELVMALDYLQN
QRIIHRDMKPDNILLDEHGHVHITDF
NIAAMLPRETQITTMAGTKPYMAPEM
FSSRKGAGYSFAVDWWSLGVTAYELL
RGRRPYHIRSSTSKEIVHTFETTVV
TYPSSAWSQEMVSLLKKLEPNPDQRF
SQLSDVQNFPMNDINWDAVFQKRLI
PGFIPNKGRLNCDPTFELEEMILESK
PLHKKKKRLAKKEKDMRKCDSSQTCL
LQEHLDSVQKEFIIFNREKVNRFN
RQPNLALEQTKDPQGED

(Pro9 to Asp390)

^ TEV protease recognition site

Tags and additions: Cleavable N-terminal His6 tag

Expression: The YANK1 protein was expressed using baculovirus infected Sf9 cells. Cells were infected at a density of 2,000,000 cells/ml for 48h.

Cell Harvest: Cells were spun at 1000x g for 20 mins and the pellets resuspended in Lysis Buffer and then frozen at -20°C.

Cell lysis: The resuspended cell pellet was thawed and lysed by sonication. PEI (polyethyleneimine) was added to a final concentration of 0.05 %. The cell debris and precipitated DNA were spun down.

Lysis buffer: 50 mM Tris pH 7.8, 200 mM NaCl, 20 mM Imidazole, 0.5 mM TCEP, 1:2000 dilution of Sigma protease inhibitor cocktail.

Column 1: 6 ml of Ni-Sepharose in a 2 cm diameter gravity flow column

Column 1 Buffers:

Binding buffer: 50 mM Hepes pH 7.4, 500 mM NaCl, 20 mM Imidazole, 0.5 mM TCEP

Wash buffer: As Binding Buffer except 1 M NaCl and 40 mM imidazole.

Elution buffer: As Binding Buffer except 60 mM imidazole.

Column 1 Procedure: The clarified supernatant was passed through the column. The column was washed with 100 ml of Binding Buffer and 50 ml of Wash Buffer 1 and 40 ml of Wash Buffer 2. 36 ml of Elute Buffer was passed through to elute the protein.

Column 2: S200 16/60 Gel Filtration (GE Healthcare)

Column 2 Buffers:

Gel Filtration buffer: 20 mM Tris pH 7.8, 150 mM NaCl, 0.5 mM TCEP

Column 2 Procedure: The eluted protein was concentrated to 5 ml volume and injected onto the column.

Concentration: The pooled fractions from the the gel filtration were passed through 0.8 ml of Ni-Sepharose (pre-equilibrated into GF Buffer) followed by 5 ml of additional GF Buffer. The resin was eluted with 5 ml of GF Buffer containing 10 mM, 20 mM and then 30 mM imidazole.

The fractions containing YANK1 were combined. Staurosporine was added and the sample concentrated to 27 mg/ml (measured by 280 nm absorbance). The sample was diluted with GF buffer and reconcentrated to reduce the imidazole concentration.

Mass spec characterization:

Expected: 45101.0

Observed: 45101.9

Crystallization: Crystals grew from a 1:2 ratio of protein and precipitant solution (0.2 M Sodium acetate, 20% PEG 3350, 10% Ethylene Glycol), using the vapour diffusion method. (A peptide was added during crystallisation, however the peptide is not visible in the structure and similar crystals also grew in the absence of peptide.)

Data Collection: Crystals were cryo-protected by equilibration into precipitant solution containing 25% ethylene glycol, and then flash frozen in liquid nitrogen. Data was collected at Diamond, beamline IO4-1.