

SETD2

PDB:4FMU

Revision

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Entry Clone Accession:GI:197313748

Entry Clone Source:MGC

SGC Clone Accession:SETD2:JMC009-H07:C43809

Tag:N-terminal: His-tag with integrated TEV protease site:MHHHHHHSSGRENLYFQG

Host:E.coli BL21 (DE3) codon plus RIL (Stratagen).

Construct

Prelude:

Sequence:

gETSVPPGSALVGPSCVMDDFRDPQRWKECAKQGKMPCYFDLIEENVYLTERKKNKSHRDIKRMQCECTPLSKDERAQGEIACGEDC
LNRLLMIECSSRCPNGDYCSNRRFQRKQHADVEVILTEKKGWGLRAAKDLPSTFVLEYCGEVLDHKEFKARVKEYARNKNIHYYFM
ALKNDEIIDATQKGNCsRFMNHSCEPNCETQKWTVNGQLRVGFFTTKLVPsGSELTfDYQfQRYGKEAQKCFcGSANCRGYLGGENR
VSIRAAGGKMKKERSRK

Vector:pET28-MHL

Growth

Medium:SETD2 was expressed in E.coli BL21(DE3) codon plus RIL in Terrific Broth (TB) medium in the presence of 50 µg/ml of kanamycin at 37°C to an OD600 of 1.5. Cells were then induced by isopropyl-1-thio-D-galactopyranoside (IPTG), final concentration 1 mM, and incubated overnight at 15°C.

Antibiotics:

Procedure:

Purification

Procedure

The crude extract was cleared by centrifugation. The clarified lysate was loaded onto 5 ml HiTrap Chelating column (Amersham Biosciences), charged with Ni²⁺. The column was washed with 10 CV of 20 mM HEPES buffer, pH 7.4, containing 250 mM NaCl and 50 mM imidazole, and the protein was eluted with elution buffer (20 mM HEPES, pH 7.4, 250 mM NaCl, 250 mM imidazole). The protein was loaded on Superdex200 column (26x60) (Amersham Biosciences), equilibrated with 20 mM PIPES buffer, pH 6.5, and 250 mM NaCl, at flow rate 4 ml/min. TEV protease was added to the pooled fractions to remove His-tag. The protein was further purified to

homogeneity by ion-exchange chromatography on Source 30S column (10x10)(Amersham Biosciences), equilibrated with buffer 20 mM PIPES, pH 6.5, and eluted with linear gradient of NaCl up to 500 mM concentration (20CV). Purification yield was 40mg of the protein per 1L of culture.

TEV cleavage.

Extraction

Procedure

Cells were harvested by centrifugation at 12,227 Xg. The cell pellets were frozen in liquid nitrogen and stored at -80°C. For the purification, 11 g of the cell paste was thawed and resuspended in 110 ml lysis buffer (50 mM HEPES, pH 7.4, 500 mM NaCl, 5 mM imidazole, 2 mM β -mercaptoethanol, 5% glycerol) with protease inhibitor (1 mM phenylmethyl sulfonyl fluoride, PMSF). The cells were lysed by passing through Microfluidizer (Microfluidics Corp.) at 20,000 psi.

Concentration: 26.1 mg/ml

Ligand

Sinefungin analog **MassSpec:** expected MW = 31902.3 Da, measured MW = 31901.6 Da.

Crystallization: Purified SETD2 (10.3 mg/ml) was complexed with Pr-SNF at 1:5 molar ratio of protein:compound and crystallized using the sitting drop vapor diffusion method at 20 °C by mixing 1 μ l of the protein solution with 1 μ l of the reservoir solution containing 20 % PEG4000, 10 % isopropanol, 0.1 M HEPES, pH 7.5.

NMR Spectroscopy:

Data Collection:

Data Processing: