

# KCNK10C

**PDB Code: 4BW5**

## Material and Methods

**Entry Clone Source:** 4BW5

**Entry Clone Accession:** IMAGE: [30915621](#)

**SGC Construct ID:** KCNK10C-c011

**GI number:** 54207

**Vector:** pFB-CT10HF-LIC

### Amplified construct sequence:

```
ATGGGCTTGCAGACCGTCATGAAGTG  
GAAGACGGTGGTGCATCTTGTGG  
TTGTGGTGGTCTACCTTGTCACTGGC  
GGTCTTGTCTCCGGCATTGGAGCA  
GCCCTTGAGAGCAGCCAGAAGAATA  
CCATCGCCTTGGAGAAGGCAGGAATT  
CTGCAGGATCATGTCGTGAGGCC  
CCAGGAGCTGGAGACGTTGATCCAGC  
ATGCTCTTGATGCTGACAATGCAGGA  
GTCAGTCCAATAGGAAACTCTTCAA  
CAACAGCAGCCACTGGGACCTCGGCA  
GTGCCTTTCTTGCTGGAAGTGT  
ATTACGACCATAGGGTATGGGAATAT  
TGCTCCGAGCACTGAAGGAGGCAAAA  
TCTTTGTATTTATATGCCATCTT  
GGAATTCCACTCTTGGTTCTTATT  
GGCTGGAATTGGAGACCAACTGGAA  
CCATCTTGGAAAAGCATTGCAAGA  
GTGGAGAAGGTCTTCGAAAAAGCA  
AGTGAGTCAGACCAAGATCCGGTCA  
TCTCAACCACCTGTTCATCTGGCC  
GGCTGCATTGTGTTGTGACGATCCC  
TGCTGTCACTTTAAGTACATCGAGG  
GCTGGACGGCCTGGAGTCCATTAC  
TTTGTGGTGGTCACTCTGACCACGGT  
GGGCTTGGTGATTGTGGCAGGGG  
GAAACGCTGGCATCAATTATCGGGAG  
TGGTATAAGCCCCTAGTGTGGTTTG  
GATCCTTGTGGCCTTGCCTACTTTG  
CAGCTGTCCTCAGTATGATCGGAGAT  
TGGCTACGGTTCTGTCCAAAAAGAC  
AAAAGAAGAGGTGGGTGAAGCAGAGA  
ACCTCTACTTCCAATCGCACCACAT  
CACCACCATCACCACCATGATTA  
CAAGGATGACGACGATAAGTGA
```

**Expressed sequence (small letters refer to tag sequence):**

MGLQTVMKWKTVAIFVVVVVYLV  
GLVFRALEQPFESSQKNTIALEKAEF  
LRDHVCSPQELETLIQHALDADNAG  
VSPIGNSSNNSSHWDLGSAFFFAGTV  
ITTIGYGNIAPISTEGGKIFCILYAI  
GIPLFGFLLAGIGDQLGTIFGKSIAR  
VEKVFRKKQVSQTKIRVISTILFILA  
GCIVFVTIPAVIFKYIEGWTalesiy  
FVVVTLTTVGFDFVAGGNAGINYRE  
WYKPLVWFWILVGLAYFAAVLSMIGD  
WLRVLSKKTKEEVGEAENLYFQ^SHH  
HHHHHHHHDYKDDDK ^ TEV cleavage site

**Tags and additions:** C-terminal, TEV cleavable deahistidine / FLAG tag.

**Host:** Spodoptera frugiperda (SF9) insect cells

**Growth medium, induction protocol:**

Insect cells with a density of 2x10<sup>6</sup> per litre of cell culture in SF900 medium (Invitrogen) were infected with recombinant baculovirus (5ml P2 virus per litre cell culture) and incubated for 65 hours. Cells were harvested by centrifugation and the pellet was flash frozen in liquid N<sub>2</sub>.

**Extraction method:** Frozen pellets were thawed and re-suspended in extraction buffer supplemented with 1% OGNG/ 0.1% CHS and incubated at 4°C on a tube rotator. The extracted protein was separated from insoluble membranes by centrifugation and collected for purification.

**Extraction buffer:**

50 mM HEPES, pH 7.5; 200 mM KCl;

**Column 1:** Co-affinity. Cobalt Talon (Clontech), 1 ml of 50 % slurry/ 1L of cells in 1.5 x 10 cm column, washed with extraction/ wash buffer.

**Buffers:**

**Wash buffer:** 50 mM HEPES, pH 7.5; 200 mM KCl; 20 mM imidazole; detergent at 3x CMC.

**Elution buffer:** 50 mM HEPES, pH 7.5; 200 mM KCl; 250 mM imidazole; detergent at 3xCMC

**Procedure:**

The solubilised membrane protein was batch bound to Cobalt Talon resin equilibrated with extraction buffer at 4°C for one hour and subsequently poured into a gravity flow glass column. The column was then washed with 30CV of wash buffer followed by elution in 1CV fractions until all the protein was eluted.

**Desalting:**

IMAC purified protein was loaded onto PD10 columns (GE healthcare) pre- equilibrated in extraction buffer supplemented with detergent at 3x CMC, and eluted with the same buffer.

**Enzymatic treatment:** TEV protease (1:10, TEV:protein) and PNGaseF (1:20, PNGaseF:protein) was added overnight at 4°C to desalting protein. The enzymes were removed and tag cleaved protein was collected by binding to Cobalt talon and collecting the flow-through.

**Column 2:** Size Exclusion Chromatography. Superose 6 (GE healthcare).

**SEC Buffer:** 20 mM HEPES pH 7.5, 200 mM KCl, detergent at 2xCMC

**Procedure:** The protein was concentrated and applied to a Superose 6 gel filtration column equilibrated with gel filtration buffer using an AKTA purifier system.

**Protein concentration:** Protein was concentrated to ~20mg/ml using a 30kDa cut-off concentrator and back diluted to 9-11mg/ml.

**Crystallization:** Crystals grew at 4°C from sitting drops (195nl) comprising 130nl of concentrated protein and 65nl of reservoir equilibrated against 20?l of reservoir solution (0.1M HEPES pH 8.0, 1mM cadmium chloride, 2% (w/v) benzamidine, 31% (v/v) PEG400).

**Cryocooling:** Crystals were cryo-cooled by slow transfer into artificial mother liquor containing 0.2 M potassium chloride, 2 % (w/v) benzamidine, 0.2 % OGNG / 0.02 % CHS, 0.1 M HEPES, pH 8.0, 35 % (v/v) PEG400 followed by plunging into liquid nitrogen. Diffraction of the crystals was improved by slowly increasing the concentration of PEG400 to above 35 % (v/v) which resulted in a 10 % shrinkage in unit cell volume compared to untreated crystals.

**Data Collection:** Data were collected at 100K on the microfocus beamline I24 at Diamond Light Source using helical / straight line scans with a beamsize of 10?m x 10?m (wxh). Diffraction was anisotropic with spots to 3.2Å in the best direction and 4.1Å in the worst direction.

**Structure Solution:** Structure was phased by molecular replacement using the coordinates for the transmembrane regions of TRAAK (3UM7) and TWIK1 (3UKM). The spacegroup is P21 with two channel homodimers per asymmetric unit. Refinement was carried out using BUSTER using NCS and TLS restraints using all data to 3.2Å.

**Resolution:** 3.4Å