

# PANK1

**PDB:3SMP**

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**BC172476

**Entry Clone Source:**Synthetic DNA from GenScript

**SGC Clone Accession:**HPC03H-A03

**Tag:**N-terminal His6-tag, not removed

**Host:**BL21-V2R

## Construct

**Prelude:**PANK1: K231-D597

**Sequence:**

```
mgsshhhhhhssglvprgsKNRPPFWGMDIGGLVKLVYFEPKDITAEEQEEVENLKSIRKYLTSNTAYGKTGIRDVHLELKNL  
TMCGRKGNLHFIRFPSCAMHRFIQMGSEKNFSSLHTTCATGGGAFKFEEDFRMIADLQLHKLDELDCLIQGLLYVDSVGFNGKPEC  
YYFENPTNPELCQKKPYCLDNYPMLLVNMGSGVSILAVYSKDNYKRVGTSLGGTFLGLCCLLTGCETFEEALEMAAKGdstNVD  
KLVKDIYGGDYERFGLQGSAVASSFGNMSKEKRDISKEDLARATLVTITNNIGSIARMCALNENIDRVVFVGNFLRINMVSML  
AYAMDFWSKGQLKALFLEHEGYFGAVGALLELFKMTDD
```

**Vector:**pET28a-LIC

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**The target was expressed in *E. coli* by inoculating 100 mL of overnight culture grown in Luria-Bertani medium into a 1.8 L of Terrific Broth medium in the presence of 50  $\mu$ g/mL kanamycin at 37°C. When OD600 was ~2.5 to 3.5, the culture was induced with 1mM IPTG and the temperature was reduced to 18°C, and the cells were allowed to grow overnight before harvesting and flash frozen by liquid nitrogen.

## Purification

### Procedure

The lysate was centrifuged at 15000 rpm for 30 min and the supernatant was passed through DE52 (Whatman) column equilibrated with the binding buffer and then loaded onto 3 mL Ni-NTA column (Qiagen) equilibrated with the same binding buffer at 4 °C. The Ni-NTA column was washed with 150 mL of the wash buffer (10mM Tris pH 7.5, 0.5 M NaCl, 5% glycerol, 30 mM imidazole) and the protein was eluted with 15 mL of the elution buffer (10mM Tris pH 7.5, 0.5 M NaCl, 5% glycerol, 250 mM imidazole). The protein were further purified and desalted

using gel filtration column, Superdex 200 (26/60), which was pre-equilibrated with 20 mM Tris pH 8.0, 0.2 M NaCl, and 10 mM DTT. All proteins were concentrated using an Amicon Ultra centrifugal filter to a final concentration of 30 mg/mL. Protein concentrations were measured using Bradford assay with purity >95% based on SDS-PAGE analysis.

## Extraction

### Procedure

The thawed cell pellets were suspended in 100 mL of the binding buffer (10 mM Tris pH 7.5, 0.5 M NaCl, 5% glycerol, 5 mM imidazole) with a protease inhibitor cocktail (0.1 mM M benzamidine-HCl and 0.1 mM phenylmethyl sulfonyl fluoride), and 0.5% CHAPS. The cells were lysed by Microfluidizer (Microfluidics M110-EH).

**Concentration:** 30 mg/mL

### Ligand

#### Acetyl Coenzyme A

**MassSpec:** Crystallization trials were set up using the sitting drop vapor diffusion method and the protein drop was equilibrated against a reservoir solution with 1:1 volume ratio. Prior to crystallization, the purified protein was incubated overnight at 4 °C in the presence of Acetyl Coenzyme A (10~15 mM final concentration). Crystals of PANK1 were grown at 20% PEG 1500, 0.2 M Magnesium Chloride, 0.1 M Sodium Cacodylate, pH 5.5.

### NMR Spectroscopy:

### Data Collection:

### Data Processing: