

# MLH1

**PDB:3RBN**

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**GI: 4557757

**Entry Clone Source:**MGC

**SGC Clone Accession:**MLH1:JMC01L-G02:C205695

**Tag:**N-terminal: His-tag with integrated TEV protease site: MHHHHHHSSGRENLYFQG

**Host:**E.coli BL21 (DE3) V2R-pRARE

## Construct

**Prelude:**

**Sequence:**

gSRKEMTAACTPRRIIINLTSVLSLQEEINEQGHEVLREMLHNHSFVGCVNPQWALAQHQTLYLLNTKLSEELFYQILIYDFANF  
GVLRLSEPAPLFDLAMLALDSPESGWTEEDGPKEGLAEYIVEFLKKKAEMPLADYFSLEIDEEGNLIGLPLLIDNYVPPLEGPIFIL  
RLATEVNWDEEKECFESLSKECAMFYSIRKQYISEESTLSGQQSEVPGSIPNSWKWTVEHIVYKALRSHILPPKHFTEGNIQLAN  
LPDLYK

**Vector:**pET28-MHL

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**MLH1 was expressed in E.coli BL21 (DE3) V2R-pRARE in M9 medium in the presence of 50 µg/ml of kanamycin. Cell were grown at 37°C to an OD600 of 1.5 and induced by isopropyl-1-thio-D-galactopyranoside (IPTG), final concentration 1 mM, in the presence of 50 mg/L of SeMet and incubated overnight at 15°C .

## Purification

**Procedure**

The lysate was loaded onto 5 ml HiTrap column (Amersham Biosciences), charged with Ni2+. The column was washed with 10 CV of 20 mM Tris-HCl, pH 8.0, containing 250 mM NaCl and 5% glycerol. The lysate was loaded onto 5 ml HiTrap column (Amersham Biosciences), charged with Ni2+. The column was washed with 10 CV of 20 mM Tris-HCl pH 8.0, containing 250 mM NaCl and 50 mM imidazole, 5% glycerol, and the protein was eluted with elution buffer (20 mM Tris-HCl pH 8.0, 250 mM NaCl, 250 mM imidazole, 5% glycerol).The protein was then loaded on to a Superdex200 (26x60, Amersham Biosciences) column equilibrated in 20 mM Tris-HCl,

pH 8.0 buffer containing 150 mM NaCl. The protein was further purified to homogeneity by ion-exchange chromatography on Source 30Q column (10x10) (Amersham Biosciences), equilibrated with buffer 20 mM Tris-HCl, pH 8.0, and eluted with linear gradient of NaCl up to 500 mM concentration (20CV). Purification yield was 23 mg of the protein per 1L of culture.

## Extraction

### Procedure

Cells were harvested by centrifugation at 7,000 rpm. The cell pellets were frozen in liquid nitrogen and stored at -80°C. For the purification the cell paste was thawed and resuspended in lysis buffer (50mM Tris pH 8.0, 0.25 M NaCl, 2 mM  $\beta$ -mercaptoethanol, 5% glycerol, 0.1% CHAPS) with protease inhibitor (1mM phenylmethyl sulfonyl fluoride, PMSF). The cells were lysed by passing through Microfluidizer (Microfluidics Corp.) at 20,000 psi.

**Concentration:** 22 mg/ml

### Ligand

**MassSpec:** Expected MW is 32996.2 Da, measured mass is 32998.3 Da.

**Crystallization:** Purified MLH1 was crystallized using sitting drop vapor diffusion method at 20 °C by mixing 1  $\mu$ l of the protein solution (8 mg/ml) with 1  $\mu$ l of the reservoir solution containing 22% PEG 4,000, 0.2 M NaOAc, 0.1 M Tris-HCl, pH 8.0.

**NMR Spectroscopy:**

**Data Collection:**

**Data Processing:**