

# PREX1

**PDB:**3QIK

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**BC053616

**Entry Clone Source:**Open Biosystems

(actual DNA used HPC0A0-B07, a chimeric full length DNA of BC053616 and synthesized DNA)

**SGC Clone Accession:**HPC0AJ-H05

**Tag:**N-terminal His6-tag, removed before crystallization

**Host:**BL21-V2R-pRARE2

## Construct

**Prelude:**PREX1:K607-A706

**Sequence:**

gKNKQLRNDFKLVENTLAKRLLTLPQEEDYGF DIEEKNKAVVVKS VQRGS LAEVAGLQVGRKIY S INEDLVFLRPFSEVESILNQSF  
CSRRPLRLLVATKA

**Vector:**pET28-MHL

## Growth

**Medium:**Terrific Broth medium in the presence of 50 mg/mL kanamycin and 25 mg/mL chloramphenicol

**Antibiotics:**

**Procedure:****LEX Bubbling.** The target protein was expressed in *E. coli* by inoculating 100 mL of overnight culture grown in Luria-Bertani medium into a 2 L of growth medium at 37 °C. When OD<sub>600</sub> reached ~3.0, the temperature of the medium was lowered to 15 °C and the culture was induced with 1 mM IPTG. The cells were allowed to grow overnight before harvested and flash frozen in liquid nitrogen and stored at -80 °C.

## Purification

### Procedure

The lysate was centrifuged at 15,000 rpm for 45 minutes and the supernatant were mixed with 4 mL 50% flurry of Ni-NTA beads and incubated at 4 degree Celsius on rotary shaker for one hour. The mixture was then centrifuged at 2300 rpm for 5 min and the supernatant discarded. The beads were then washed with 50 mL binding buffer containing 5 mM Imidazole twice. Bound proteins were eluted using 15 mL elution buffer. The flow-through was collected and further

purified by a Superdex-75 gel filtration column pre-equilibrated with gel filtration buffer. Fractions containing the target protein were pooled and concentrated using Amicon Ultra-15 centrifugal filter (mwco 10 kDa). The purity of the preparation is tested by SDS-PAGE to be greater than 95%.

## Extraction

### Procedure

Frozen cells from 4L TB culture were thawed and resuspended in 5 mL extraction buffer per gram of cell pellets with freshly added 0.5% CHAPS, and supplemented with protease inhibitor cocktail (SIGMA Catalog # P8849), and 3 uL benzonase (Sigma Catalog # E1014, 250U/uL), and lysed using sonication for 5 min at 100 W, 10 sec on/10 sec off duty cycle.

**Concentration:** 6.9 mg/mL

### Ligand

**MassSpec:** Uncut version native protein expected 13638.7, measured 13639.0

Cut version, expected 11502.4, measured 11494.5, not sure about the cause of -7.9 difference

**Crystallization:** Crystal was initially obtained from SGC-I screen condition B12.

Crystal used for structure refinement was grown in 1.2 M Li<sub>2</sub>SO<sub>4</sub>, 0.5 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.1 M Sodium Citrate pH 6.2 in sitting drop setup, using 1 uL protein + 1 uL well solution against 1 mL reservoir buffer at room temperature. Crystal used for phasing was Pt derivative of optimized crystal grown in 0.5 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 1.2 M Li<sub>2</sub>SO<sub>4</sub>, 0.1 M Sodium Citrate pH 5.6. Crystal was harvested from original drop and soaked in 4 uL of the mother liquid containing 1 mM K<sub>2</sub>PtI<sub>6</sub> for 6 hours and harvested for diffraction test.

Crystals grow to a mountable size within one week.

Cryo used paratone-N

### NMR Spectroscopy:

### Data Collection:

### Data Processing: