

# GIMAP1

**PDB:**3LXW

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**AT72-F5:BC040736.1

**Entry Clone Source:**MGCHPC09G-H01

**SGC Clone Accession:**mhhhhhssgrenlyfq\*g

**Tag:**GIMAP1:E25-R253

Tag not removed

**Host:**BL21-V2R-pRARE2

## Construct

**Prelude:**

**Sequence:**

mhhhhhssgrenlyfqgESTRRLILVGRTGAGKSATGNSILGQRRFFSRLGATSVTRACTTGSRRWDKCHVEVVDTPDIFSSQVSK  
TDPGCEERGH CYLLSAPGPHALLVTQLGRFTAQDQQAVRQVRDMFGEDVLKWMVIVFTRKEDLAGGSLHDYVSN TENRALRELVAE  
CGGRVCAFDNRATGREQEAQVVQLLGMVEGLVLEHKGAHYSNEVYELAQVLRWAGPEERLRRVAERVAARVQR

**Vector:**pET28-MHL

## Growth

**Medium:**Terrific Broth medium in the presence of 50 mg/mL kanamycin and 25 mg/mL chloramphenicol

**Antibiotics:**

**Procedure:LEX Bubbling.** The target protein was expressed in *E. coli* by inoculating 100 mL of overnight culture grown in Luria-Bertani medium into a 2 L of growth medium at 37 °C. When OD<sub>600</sub> reached ~3.0, the temperature of the medium was lowered to 15 °C and the culture was induced with 1 mM IPTG. The cells were allowed to grow overnight before harvested and flash frozen in liquid nitrogen and stored at -80 °C.

## Purification

### Procedure

The lysate was centrifuged at 15,000 rpm for 45 minutes and the supernatant were mixed with 5 mL 50% slurry of Talon Cobalt beads and incubated at 4 degrees Celsius on rotary shaker for one hour. The mixture was then centrifuged at 2300 rpm for 5 min and the supernatant discarded. The beads were then washed with washing buffer containing 30 mM and 75 mM Imidazole, and finally the elution buffer. The flow-through was collected and treated with TEV protease for two days. The protease and uncut protein were removed by flowing the solution through a Talon cobalt open column and the flow-through were further purified by a Superdex-75 gel filtration column pre-equilibrated with gel filtration buffer. Fractions containing the protein were collected and added 5mM MgCl<sub>2</sub> and concentrated with Amicon Ultra-15 centrifugal filter. GDP was then added to the concentrated protein solution to a final concentration of 5 mM. The purity of the preparation is tested by SDS-PAGE to be greater than 95%.

## Extraction

### Procedure

Frozen cells from 2L TB culture were thawed and resuspended in 150 mL extraction buffer with freshly added 0.5% CHAPS, and supplemented with protease inhibitor cocktail (SIGMA Catalog # P8849), and 3 uL benzonase (Sigma Catalog # E1014, 250U/uL), and lysed using microfluidizer at 15,000 PSI.

**Concentration:** 29.9 mg/mL

### Ligand

GDP, Mg<sup>2+</sup> **MassSpec:** Native protein expected: 27705.22 (uncut), 25568.9

Measured: 27688.1 (uncut), 25552.0

A delta of -17 Da prompts a mutation from Asn to Pro but this needs to be confirmed by DNA sequencing.

**Crystallization:** Crystal used for structure determination was grown in Red Wings screen condition E10 from initial screen.

Crystal was grown in 25% W/V PEG3350, 0.2 M NaCl, 0.1 M HEPES pH 7.5, in the presence of 1:100 (w/w) dispase and 5mM GDP in sitting drop setup.

Crystals grow to a mountable size within 3 days.

Cryoprotectant used Paratone-N

### NMR Spectroscopy:

#### Data Collection:

#### Data Processing: