

<b>Entry Clone Source:</b> Collaborator
<b>Entry Clone Accession:</b> n/a
<b>SGC Construct ID:</b> SDHLA-c019
<b>GenBank GI number:</b> gi 33694276
<b>Vector:</b> pET28a. Details [ <a href="#">PDF</a> ]; Sequence [ <a href="#">FASTA</a> ] or [ <a href="#">GenBank</a> ]
<b>Host:</b> BL21 (DE3) pLysE
<b>Tags and additions:</b> N-terminal non-cleavable His-tag
<p><b>Expressed sequence (vector-incorporated sequence in lowercase):</b></p> <p>mgsshhhhhhssglvprgshmasmtggqq  mrgsaMLPNTGRLAGCTVFITGASRGIG  KAIALKAAKDGANIVIAAKTAQPHPKLLG  TIYTAAEEIEAVGGKALPCIVDVRDEQQI  SAAVEKAIAKKFGGIDILVNNASAI SLTNT  LDTPTKRLDLMMNVNTRGTYLASKACIPY  LKKSQVAHILNISPPNLNPVWFKQHCAY  TIAKYGMSMYVLGMAEEFKGEIAVNALWP  KTAIHTAAMDMLGGPGIESQCRKVDI IAD  AAYSIFQKPKSFTGNFVIDENILKEEGIE  NFDVYAIKPGHPLQPDFFLDEYPEAVSKK  VESTGAVPElacgrtrappppplrsgc</p>
<p><b>Expression:</b> BL21 glycerol stock harbouring the SDHLA-pET28a plasmid was inoculated into 50ml TB medium supplemented with 100µg/ml ampicillin and 34µg/ml chloramphenicol and grown overnight at 37°C, 200 rpm. 10ml overnight culture was inoculated into 4x 1L TB supplemented with 100µg/ml ampicillin and incubated at 37°C, 160rpm. At OD<sub>600</sub> ~1.0, 0.2mM IPTG was added and the culture was incubated overnight at 18°C, 160rpm. The following morning cells were harvested by centrifuging for 10min at 6000rpm, and were resuspended in 70ml lysis buffer.</p>
<p><b>Extraction:</b> Cells were lysed by sonication, frozen and then re-thawed. The lysate was centrifuged at 12,000rpm for 20min.</p>
<p><b>Buffers:</b></p> <p><b>Lysis Buffer:</b> 20 mM Tris pH 8.0, 100 mM NaCl, 20% Glycerol, 50mM Imidazole, 1mM PMSF;</p> <p><b>Wash Buffer:</b> 20 mM Tris pH 8.0, 800 mM NaCl, 20% Glycerol, 50mM Imidazole, 1mM PMSF;</p> <p><b>Elution buffer:</b> 20 mM Tris pH 8.0, 500 mM NaCl, 20% Glycerol, 50mM Imidazole, 1mM PMSF;</p> <p><b>Dialysis buffer:</b> 20 mM Tris pH 8.0, 500 mM NaCl, 10% Glycerol, 1mM TCEP</p>
<p><b>Procedure:</b> The clarified lysate was adjusted to [NaCl] ~800mM, incubated with 0.5 ml of Ni-NTA resin for 25 min and then loaded onto a gravity flow column. Ni-NTA was washed with 20ml wash buffer. Elution buffer was then added to the column and protein was collected in 5 ml fractions which were analyzed by SDS-PAGE. The pooled fractions containing the protein were then dialyzed into dialysis buffer overnight at 4°C.</p>



**Concentration:** Using Amicon Ultra-15 concentrators with 10kDa cutoff the protein was concentrated to 5.3 mg/ml.

**Mass specrometry characterization:** The calculated mass of the tagged protein (36,999Da) was experimentally confirmed by mass spec analysis.

**Crystallization with NADP and 17 $\beta$ -Estradiol:** Prior to crystallization protein was supplemented with 1.5 mM NADP and 0.75 mM 17 $\beta$ -estradiol. Crystals were grown at 20°C in 150nl sitting drops mixing SDHLA (5.3 mg/ml) with reservoir solution containing 0.1 M BIS-TRIS pH 5.5, 25% PEG 3350 in a 2:1 ratio. Crystals were cryo-protected with 25% glycerol before flash-freezing in liquid nitrogen.

**Data collection: Resolution:** 2.25 Å; **Beamline:** Diamond Light Source beamline I24