

FXR1

PDB:3KUF

Revision

Revision Type:created

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Entry Clone Accession:BC028983

Entry Clone Source:MGC: AT42-B4

SGC Clone Accession:

Tag:N-terminal hexahistidine tag with integrated TEV protease cleavage site:
mhhhhhssgrenlyfq*g.

Host:*E. coli* BL21(DE3)-V2R-pRARE2

Construct

Prelude:

Sequence:

mhhhhhssgrenlyfqgAELTVEVRGSGAFYKGFIKDVHEDSLTVVFENNWQPERQVPFNEVRLPPPPDIKKEISEGDEVEVYSR
ANDQEPGWWLAKVRMMKGFEFYVIEYAACDATYNEIVTFERLRPVNQKTVKKNTFFKCTVD

Vector:pET28-MHL

Growth

Medium:M9 SeMET growth media (Medicilon Inc).

Antibiotics:

Procedure:A fresh transformation was used to inoculate 20 mL LB media containing 50 µg/mL kanamycin and 30 µg/mL chloramphenicol . The culture was grown overnight at 37°C with shaking. The next day this starter culture was used to innoculate 2L of M9 SeMET growth medium. The culture was grown in LEX at 37°C to OD600 of 2.3. Methionine biosynthesis inhibition and IPTG-based induction were carried out according to the manufacturer's protocol. The temperature was reduced to 14°C and the culture was incubated for a further 18 hours before harvesting the cells.

Purification

Procedure

Column 1: Affinity purification.

open Ni-NTA column Procedure: The supernatant was incubated with 6mL of 50% slurry Ni-NTA beads by rocking. After 1 hour incubation at 4°C, the beads were washed with 50 mL of lysis buffer. The protein was eluted using ~20mL EB.

Column 2: Size Exclusion, HiLoad 16/60 Superdex 75

Prep Grade Procedure: The eluent from the NiNTA column was concentrated and loaded onto the size exclusion column at 1 mL/min, fraction size 2mL. The fractions containing protein were identified on a SDS-PAGE gel.

Extraction**Procedure**

Cells were harvested by centrifugation and pellets were stored in -80°C. Prior to purification, the cell pellet was resuspended in lysis buffer. Cells were disrupted by sonication (10 minutes) and samples were centrifuged for 60 min at 70000 g.

Concentration: 10 mg/ml.

Ligand**MassSpec:**

Crystallization: 1.4M (NH₄)₂SO₄, 0.1 M HEPES pH 6.8, 0.25M NaCl, 10 mM DTT by sitting-drop vapour diffusion, using micro seeding after equilibration for 3 hours. The drop was prepared by mixing 1 microL protein with 1 microL of reservoir solution. Protein concentration is 7.25mg/ml. Crystals appear after a period of 2 days.

NMR Spectroscopy:

Data Collection: Crystals were cryo-protected using mother liquor containing 20% glycerol, and flash frozen in liquid nitrogen. Diffraction data were collected APS 19-ID to 2.75 Å.

Data Processing: