

Entry Clone Source: Site-directed mutagenesis Thr113Ala (<u>underlined below</u>)
Entry Clone Accession: n/a
SGC Construct ID: UGDHA-c704
GenBank GI number: gi 4507813
Vector: pBEN1-SGC. Details [PDF]; Sequence [FASTA] or [GenBank]
<p>Amplified construct sequence:</p> <pre> CATATGGACCCCGAAGAGGCGAGTGTTACT TCAACAGAAGAAACCTTAACGCCAGCACAG GAGGCCGCACGCACCCGCGCTGCTAACAAA GCCCCGAAAGGAAGCTGAGTTGGCTGCTGCC ACCGCTGAACAACTAGTGACGAGAAGACC ACCGGCTGGCGGGGCGGCCACGTGGTGGAG GGCCTGGCCGGCGAGCTGGAGCAGCTGCGG GCCAGGCTGGAGCACCACCCTCAGGGCCAG CGGGAGCCCTCCGGCGGCTGCAAGCTGGGC CTGGGTACCGAGAACCTGTACTTCCAATCC ATGTTTGAAATTAAGAAGATCTGTTGCATT GGTGCAGGCTATGTTGGAGGACCCACATGT AGTGTCAATTGCTCATATGTGTCCTGAAATC AGGGTAACGGTTGTTGATGTCAATGAATCA AGAATCAATGCGTGGAATTCTCCTACACTT CCTATTTATGAGCCAGGACTAAAAGAAGTG GTAGAATCCTGTGAGGAAAAAATCTTTTT TTTTCTACCAATATTGATGATGCCATCAAA GAAGCTGATCTTGTATTTATTTCTGTGAAT ACTCCAACAAAAACCTATGGAATGGGGAAA GGCCGGGCAGCAGATCTGAAGTATATTGAA GCTTGTGCTAGACGCATTGTGCAAAACTCA AATGGGTACAAAATTGTGACTGAGAAAAGC GCAGTTCCGGTGCGGGCAGCAGAAAGTATC CGTCGCATATTTGATGCAAAACACAAAACCC AAGTTGAATTTACAGGTGCTGTCCAACCCT GAGTTTCTGGCAGAGGGAACAGCCATCAAG GACCTAAAGAACCCAGACAGAGTACTGATT GGAGGGGATGAAACTCCAGAGGGCCAGAGA GCTGTGCAGGCCCTGTGTGCTGTATATGAG CACTGGGTTCCAGAGAAAAGATCCTCACC ACTAATACTTGGTCTTCAGAGCTTTCCAAA CTGGCAGCAAATGCTTTTCTTGCCCAGAGA ATAAGCAGCATTAACTCCATAAGTGCTCTG TGTGAAGCAACAGGAGCTGATGTAGAAGAG GTAGCAACAGCGATTGGAATGGACCAGAGA ATTGGAAACAAGTTTCTAAAAGCCAGTGTT GGGTTTGGTGGGAGCTGTTTCCAAAAGGAT GTTCTGAATTTGGTTTATCTCTGTGAGGCT CTGAATTTGCCAGAAGTAGCTCGTTATTGG CAGCAGGTCATAGACATGAATGACTACCAG AGGAGGAGGTTTGCTTCCCGGATCATAGAT AGTCTGTTTAATACAGTAACTGATAAGAAG ATAGCTATTTGGGATTTGCATTCAAAAAG GACACTGGTGATACAAGAGAATCTTCTAGT ATATATATTAGCAAATATTTGATGGATGAA GGTGCACATCTACATATATATGATCCAAA GTACCTAGGGAACAAATAGTTGTGGATCTT </pre>

TCTCATCCAGGTGTTTCAGAGGATGACCAA
GTGTCCCGGCTCGTGACCATTTCCAAGGAT
CCATATGAAGCATGTGATGGTGCCCATGCT
GTTGTTATTTGCACTGAGTGGGACATGTTT
AAGGAATTGGATTATGAACGCATTCATAAA
AAAATGCTAAAGCCAGCCTTTATCTTCGAT
GGACGGCGTGTCCTGGATGGGCTCCACAAT
GAACTACAAACCATTGGCTTCCAGATTGAA
ACAATTGGCAAAAAGGTGTGACAGTAAAGG
TGGATACGGATCCGAATTCGAGCTCCGTCG
ACAAGCTTGCGGCCGCACTCGAGCACCACC
ACCACCACCACTGA

Final protein sequence (tag sequence in lowercase):

mdpeeasvtsteetltpageaartraanka
rkeaelaaataeqltsdekttgwrgghvveg
lageleqlrarlehqpqqrepssggcklgl
gtenlyfq*sMFEIKKICIGAGYVGGPTC
SVIAHMCPEIRVTVDVNESRINAWNSPTL
PIYEPGLKEVVESCRGKNLFFSTNIDDAIK
EADLVFISVNTPTKTYGMGKGRAADLKYIE
ACARRIVQNSNGYKIVTEKS₄VPVRAAESI
RRIFDANTKPNLNLQVLSNPEFLAEGTAIK
DLKNPDRVLIGGDETPEGQRAVQALCAVYE
HWVPREKILTNTWSSSELSKLAANAFQAQR
ISSINSISALCEATGADVEEVATAIGMDQR
IGNKFLKASVGGFGSCFQKDVNLVYLCEA
LNLPEVARYWQQVIDMNDYQRRRFASRIID
SLFNTVTDKKIALGFAFKKDTGDTRESSS
IYISKYLMDEGAHLHIYDPKVPREQIVVDL
SHPGVSEDDQVSRVLTISKDPYEACDGAHA
VVICTEWD MFKELDYERIHKKMLKPAFIFD
GRRVLDGLHNELQTIGFQIETIGKKV

Tags and additions: N-terminal SET1 and SBP tags, followed by a TEV protease cleavage site:

mdpeeasvtsteetltpageaartraanka
rkeaelaaataeqltsdekttgwrgghvveg
lageleqlrarlehqpqqrepssggcklgl
gtenlyfq*s (* - TEV cleavage site)

Tag removed: yes

Host: BL21(DE3)-R3-pRARE2

Expression protocol: A glycerol stock of host strain BL21(DE3)-R3-pRARE2 carrying the expression plasmid was used to inoculate 10 ml of TB (terrific Broth) supplemented with 50 µg/ml kanamycin and 34 µg/ml chloramphenicol. This starter culture was grown overnight at 37°C and used to inoculate 6X 1 liter culture in the same media (initial OD₆₀₀ = 0.01). The culture was grown at 37°C until the OD₆₀₀ reached ~0.8. After that the temperature was lowered to 18°C and protein production was induced with 0.2 mM IPTG. Recombinant UGDHA was expressed at that temperature overnight. The next day cells were harvested by centrifugation at 4000 rpm for 30 minutes and the pellet was resuspended in 150 ml Streptavidin binding buffer supplemented with Complete Protease Inhibitors (1 tablet/50 ml) and stored at -20°C.

Cell extraction:

Streptavidin binding buffer: 20 mM Tris, pH 8.0, 150 mM NaCl.

Procedure: Frozen cell pellets were thawed and lysed by passing 5 times through a high pressure homogenizer. The lysate was clarified by centrifuging for 60 minutes at 21,000 rpm at 4°C. Before applying to the column the lysate was further clarified by passing through a 1.2µm syringe filter

Column 1: Streptavidin sepharose

Column 1 Buffers: Binding and washing buffer: 20 mM Tris, pH 8.0, 150 mM NaCl; Elution buffer: 20 mM Tris/HCl, pH 8.0, 150 mM NaCl, 2mM Biotin.
Column 1 Procedure: Ten ml Streptavidin sepharose resin was equilibrated with 100 mL of binding buffer. The supernatant was incubated with the washed resin at 4°C for 2 hour by gentle rotation and packed into the column. After the lysate had passed through the column it was washed with 60 ml of washing buffer. The protein was eluted with 30 ml of elution buffer. The eluted protein was concentrated (Vivaspin centricon MWCO 30kDa) and exchanged into gel filtration buffer using a PD-10 desalting column.
Enzymatic treatment : His-tagged TEV protease was added to the protein using a 1:20 TEV to protein ratio (mg/mg). The digestion was incubated 48 hours at 4°C.
Column 2: Gel filtration, Hiload 16/60 Superdex S200 prep grade, 120 ml (GE Healthcare)
Column 2 Buffer: Gel Filtration Buffer - 50mM HEPES pH 7.5, 300mM NaCl, 0.5 mM TCEP
Column 2 Procedure: Following TEV digestion, the protein sample was loaded onto an S200 column at 1 ml/min using an AKTA Purifier system at 4°C. Fractions were analysed by SDS - PAGE and pooled according to purity. The eluted protein was concentrated (Vivaspin centricon MWCO 30kDa) and buffer exchanged into ion exchange buffer A using a PD-10 desalting column.
Column 3: MonoQ 5/50 (Qiagen)
Column 3 Buffers: Buffer A: 50 mM HEPES, pH 7.5; Buffer B: 50 mM HEPES, pH 7.5, 1 M NaCl
Column 3 Procedure: The protein was applied to the column in Buffer A and eluted using a linear gradient from 0 - 50% Buffer B. After SDS-Page analysis of the fractions, pure protein was pooled and concentrated using a Vivaspin centricon with a MWCO of 30kDa.
Mass spectrometry characterization: ESI-MS revealed that the protein had a mass of 51991 Da (Expected mass 51990 Da).
Crystallization: Crystals were grown at 20°C by vapour diffusion in sitting drops by mixing protein (20 mg/ml) and well solution containing 15% PEG smear, 0.1M MES, pH 6.0 at a protein to precipitant ratio of 1:1. A crystal was cryo-protected using well solution supplemented with 20% (v/v) ethylene glycol and flash-cooled in liquid nitrogen.
Data Collection: Resolution (scaled): 2.4 Å X-ray source: Diamond light source I03.