

**Entry Clone Source:** Site-directed mutagenesis Thr113Ala (underlined below)

**Entry Clone Accession:** n/a

**SGC Construct ID:** UGDHA-c704

**GenBank GI number:** gi|4507813

**Vector:** pBEN1-SGC. Details [[PDF](#)]; Sequence [[FASTA](#)] or [[GenBank](#)]

**Amplified construct sequence:**

CATATGGACCCCGAAGAGGGCGAGTGTTACT  
TCAACAGAAGAACCTTAACGCCAGCACAG  
GAGGCCGCACGCACCCGCGCTGCTAACAAA  
GCCCGAAAGGAAGCTGAGTTGGCTGCTGCC  
ACCGCTGAACAAACTAGTGACGAGAAGACC  
ACCGGCTGGCGGGCGGCCACGTGGTGGAG  
GCCCTGGCCGGCGAGCTGGAGCAGCTGCC  
GCCAGGCTGGAGCACCACCCCTAGGGCCAG  
CGGGAGCCCTCCGGCGCTGCAAGCTGGC  
CTGGGTACCGAGAACCTGTACTTCCAATCC  
ATGTTGAAATTAAAGAAGATCTGTTGCATT  
GGTGCAGGCTATGTTGGAGGACCCACATGT  
AGTGTCTTGTCTATGTGTCTGAAATC  
AGGGTAACGGTTGTTGATGTCAATGAATCA  
AGAATCAATGCGTGGATTCTCCTACACTT  
CCTATTATGAGCCAGGACTAAAAGAAGTG  
GTAGAATCCTGTCGAGGAAAAATCTTTT  
TTTCTACCAATATTGATGATGCCATCAA  
GAAGCTGATCTGTATTATTCTGTGAAT  
ACTCCAACAAAAACCTATGGAATGGGGAAA  
GGCCGGGCAGCAGATCTGAAGTATATTGAA  
GCTTGTGCTAGACGCATTGTGCAAAACTCA  
AATGGGTACAAAATTGTGACTGAGAAAAGC  
GCAGTTCCGGTGCAGCAGCAGAAAGTATC  
CGTCGCATATTGATGCAAACACAAAACCC  
AACCTGAATTACAGGTGCTGTCCAACCC  
GAGTTTCTGGCAGAGGGAACAGCCATCAAG  
GACCTAAAGAACCCAGACAGAGTACTGATT  
GGAGGGGATGAAACTCCAGAGGGCCAGAGA  
GCTGTGCAGGCCCTGTTGCTGTATATGAG  
CACTGGGTTCCCAGAGAAAAGATCCTCACC  
ACTAATACTGGTCTTCAGAGCTTCCAAA  
CTGGCAGCAAATGCTTTCTGCCAGAGA  
ATAAGCAGCATTAACCCATAAGTGCTCTG  
TGTGAAGCAACAGGAGCTGATGAGAG  
GTAGCAACAGCGATTGGAATGGACCAGAGA  
ATTGGAAACAAGTTCTAAAAGCCAGTGTT  
GGGTTTGGTGGGAGCTGTTCCAAAAGGAT  
GTTCTGAATTGGTTATCTCTGTGAGGCT  
CTGAATTGCCAGAAGTAGCTCGTTATTGG  
CAGCAGGTCATAGACATGAATGACTACCAG  
AGGAGGAGGTTGCTCCGGATCATAGAT  
AGTCTGTTAATACAGTAACTGATAAGAAG  
ATAGCTATTGGGATTGCAATTCAAAAAG  
GACACTGGTGATACAAGAGAATCTCTAGT  
ATATATATTAGCAAATATTGATGGATGAA  
GGTGCACATCTACATATATGATCCAAA  
GTACCTAGGAAACAAATAGTTGTGGATCTT

TCTCATCCAGGTGTTTCAGAGGATGACCAA  
GTGTCCCGGCTCGTGACCATTCCAAGGAT  
CCATATGAAGCATGTGATGGTGCCCAGTGC  
GTGTTATTGCACTGAGTGGGACATGTT  
AAGGAATTGGATTATGAACGCATTATAAA  
AAAATGCTAAAGCCAGCCTTATCTCGAT  
GGACGGCGTGTCTGGATGGGCTCCACAAAT  
GAACTACAAACCATTGGCTCAGATTGAA  
ACAATTGGCAAAAGGTGTGACAGTAAAGG  
TGGATACGGATCCGAATTGAGCTCCGTCG  
ACAAGCTTGGCCGCACTCGAGCACCACC  
ACCACCACTGA

**Final protein sequence (tag sequence in lowercase):**

mdpeasvtsteetltpaqeartraanka  
rkeaelaaataeqtsdekttgwrgghvveg  
lageleqlrarlehhpqgqrepsggcklgl  
gtenlyfq\*sMFEIKKICCIAGAYVGGPTC  
SVIAHMCPEIRVTVDVNESRINAWSPTL  
PIYEPGLKEVVESCRGKNLFFSTNIDDAIK  
EADL VFISVNTPTKTYGMGKGRAADLKYE  
ACARRIVQNSNGYKIVTEKS~~A~~VPVRAAESI  
RRIFDANTKPNLNLQVLSNPEFLAEGTAIK  
DLKNPDRVLIGGDETPEGQRAVQALCAVYE  
HWVPREKILTTNTWSSSELSKLAANAFLAQR  
ISSINSISALCEATGADVEEVATAIGMDQR  
IGNKFLKASVGFGGSCFQKDVLNLVYLCEA  
LNLPFVARYWQQVIDMNDYQRRRFASRIID  
SLFNTVTDKKIAILGFAFKKDGTRESSS  
IYISKYLMDEGAHLHIYDPKVPREQIVVDL  
SHPGVSEDDQVSRLVTISKDPYEACDGAHA  
VVICTEWDMFKELDYERIHKMLKPAFIFD  
GRRVLDGLHNELOQTIGFQIETIGKKV

**Tags and additions:** N-terminal SET1 and SBP tags, followed by a TEV protease cleavage site:

mdpeasvtsteetltpaqeartraanka  
rkeaelaaataeqtsdekttgwrgghvveg  
lageleqlrarlehhpqgqrepsggcklgl  
gtenlyfq\*s (\* - TEV cleavage site)

**Tag removed:** yes

**Host:** BL21(DE3)-R3-pRARE2

**Expression protocol:** A glycerol stock of host strain BL21(DE3)-R3-pRARE2 carrying the expression plasmid was used to inoculate 10 ml of TB (terrific Broth) supplemented with 50 µg/ml kanamycin and 34 µg/ml chloramphenicol. This starter culture was grown overnight at 37°C and used to inoculate 6X 1 liter culture in the same media (initial OD<sub>600</sub> = 0.01). The culture was grown at 37°C until the OD<sub>600</sub> reached ~0.8. After that the temperature was lowered to 18°C and protein production was induced with 0.2 mM IPTG. Recombinant UGDHA was expressed at that temperature overnight. The next day cells were harvested by centrifugation at 4000 rpm for 30 minutes and the pellet was resuspended in 150 ml Strepavidin binding buffer supplemented with Complete Protease Inhibitors (1 tablet/50 ml) and stored at -20°C.

**Cell extraction:**

**Strepavidin binding buffer:** 20 mM Tris, pH 8.0, 150 mM NaCl.

**Procedure:** Frozen cell pellets were thawed and lysed by passing 5 times through a high pressure homogenizer. The lysate was clarified by centrifuging for 60 minutes at 21,000 rpm at 4C. Before applying to the column the lysate was further clarified by passing through a 1.2µm syringe filter

**Column 1:** Streptavidin sepharose

**Column 1 Buffers: Binding and washing buffer:** 20 mM Tris, pH 8.0, 150 mM NaCl; **Elution buffer:** 20 mM Tris/HCl, pH 8.0, 150 mM NaCl, 2mM Biotin.

**Column 1 Procedure:** Ten ml Streptavidin sepharose resin was equilibrated with 100 mL of binding buffer. The supernatant was incubated with the washed resin at 4°C for 2 hour by gentle rotation and packed into the column. After the lysate had passed through the column it was washed with 60 ml of washing buffer. The protein was eluted with 30 ml of elution buffer. The eluted protein was concentrated (Vivaspin centricon MWCO 30kDa) and exchanged into gel filtration buffer using a PD-10 desalting column.

**Enzymatic treatment :** His-tagged TEV protease was added to the protein using a 1:20 TEV to protein ratio (mg/mg). The digestion was incubated 48 hours at 4°C.

**Column 2:** Gel filtration, Hiload 16/60 Superdex S200 prep grade, 120 ml (GE Healthcare)

**Column 2 Buffer: Gel Filtration Buffer** - 50mM HEPES pH 7.5, 300mM NaCl, 0.5 mM TCEP

**Column 2 Procedure:** Following TEV digestion, the protein sample was loaded onto an S200 column at 1 ml/min using an AKTA Purifier system at 4°C. Fractions were analysed by SDS - PAGE and pooled according to purity. The eluted protein was concentrated (Vivaspin centricon MWCO 30kDa) and buffer exchanged into ion exchange buffer A using a PD-10 desalting column.

**Column 3:** MonoQ 5/50 (Qiagen)

**Column 3 Buffers: Buffer A:** 50 mM HEPES, pH 7.5; **Buffer B:** 50 mM HEPES, pH 7.5, 1 M NaCl

**Column 3 Procedure:** The protein was applied to the column in Buffer A and eluted using a linear gradient from 0 - 50% Buffer B. After SDS-Page analysis of the fractions, pure protein was pooled and concentrated using a Vivaspin centricon with a MWCO of 30kDa.

**Mass spectrometry characterization:** ESI-MS revealed that the protein had a mass of 51991 Da (Expected mass 51990 Da).

**Crystallization:** Crystals were grown at 20°C by vapour diffusion in sitting drops by mixing protein (20 mg/ml) and well solution containing 15% PEG smear, 0.1M MES, pH 6.0 at a protein to precipitant ratio of 1:1. A crystal was cryo-protected using well solution supplemented with 20% (v/v) ethylene glycol and flash-cooled in liquid nitrogen.

**Data Collection:**

**Resolution (scaled):** 2.4 Å

**X-ray source:** Diamond light source I03.