

**Entry Clone Source:** MGC

**Entry Clone Accession:** IMAGE:5087393

**SGC Construct ID:** HPDA-c103

**GenBank GI number:** gi|4504477

**Vector:** Vector: pNIC-CTHF. Details [[PDF](#)] ; Sequence [[FASTA](#)] or [[GenBank](#)]

**Amplified construct sequence:**

CTTAAGAAGGAGATATACTATGGGGCAAA  
GCCTGAGAGAGGCCGATTCTCCACTCCA  
CTCTGTGACCTTCTGGTTGGCAACGCCAA  
GCAGGCCGCGTCATTCTACTGCAGCAAGAT  
GGGCTTGAACCTCTAGCCTACAGGGCCT  
GGAGACCGGTTCCCAGGAGGTGGTCAGCCA  
TGTAAATCAAACAAGGGAAGATTGTGTTGT  
CCTCTCCTCAGCGCTCAACCCCTGGAACAA  
AGAGATGGCGATCACCTGGTAAACACGG  
TGACGGAGTGAAGGACATTGCGTTGAGGT  
GGAAGAGTTGTGACTACATCGTGCAGAAAGC  
ACGGGAACGGGGCGC AAAATCATGCGGGA  
GCCCTGGGTAGAGCAAGACAAGTTGGAA  
GGTGAAGTTGCTGTGCTGCAGACGTATGG  
GGACACCACACACACCTGGTGGAGAAGAT  
GAACATACATCGGCCAATTCTTGCCTGGATA  
TGAGGCCCCAGCGTTCATGGACCCCCCTACT  
TCCTAAACTGCCAATGCACTGGAGAT  
GATCGACCACATTGTGGAAACCAGCCTGA  
TCAGGAGATGGTGTCCGCCTCCGAATGGTA  
CCTGAAAAAACCTGCAGTTCCACCGCTTCTG  
GTCCTGGATGACACGCAGGTGCACACCGA  
ATATAGCTCTGCGATCCATTGTGGTGGC  
CAACTATGAAGAGTCATCAAGATGCCAT  
CAATGAGCCAGCGCCTGGCAAGAAGAAGTC  
CCAGATCCAGGAATATGTGGACTATAACGG  
GGCGCTGGGTCCAGCACATCGCTCTCAA  
GACCGAAGACATCATCACAGCGATTGCCA  
CTTGAGAGAGAGAGGGCTGGAGTTCTTATC  
TGTTCCTCCACGTACTACAAACAATGCG  
GGAGAAGCTGAAGACGGCCAAGATCAAGGT  
GAAGGAGAACATTGATGCCCTGGAGGAGCT  
GAAAATCCTGGTGGACTACGACGAGAAAGG  
CTACCTCCTGCAGATCTCACCAAACCGGT  
GCAGGACCGGCCACCGCTCTGGAAAGT  
CATCCAGCGCCACAACCACCAGGGTTTGG  
AGCCGGCAACTCAACTCACTGTTCAAGGC  
TTTCGAGGGAGGAGCAGAACCTGCGGGGTAA  
CCTCACCAACATGGAGACCAATGGGTGGT  
GCCCGGCATGGCAGAGAACCTACTTCCA  
ATCGCACCACATCACACCACCATGATTACAA  
GGATGACGACGATAAGTGAGGATCC

**Final protein sequence (tag sequence in lowercase):**

MGAKPERGRFLHFHSVT FWVGNAKQASFY  
CSKMGFEPPLAYRGLETGSREVVSHVIKQGK  
IVFVLSSALNPWNKEMGDHLVKHGDGVKDI  
AFEVEDCDYIVQKARERGAKIMREPWVEQD

KFGKVKFAVLQTYGDTTHTLVEKMNYIGQF  
LPGYEAPAFMDPLPKLPKCSLEMIIDHIVG  
NQPDQEMVSASEWYLNQFHRFWSVDDTQ  
VHTEYSSLRSIVVANYEESIKMPINEPAPG  
KKKSQIQEYVDYNGGAGVQHIALKTEDIIT  
AIRHLRERGLEFLSVPSTYYKQLREKLKTA  
KIKVKENIDALEELKILVDYDEKGYLLQIF  
TKPVQDRPTLFLEVIQRHNNQGFGAGNFNS  
LFKAFEEEQNLRGNLTNMETNGVPGMaen  
lyfq(\*)shhhhhdykddddk

Residues **aenlyfq** originate from the vector and remain after the TEV cleavage of the hexahistidine tag.

**Tags and additions:** C-terminal, TEV cleavable hexahistidine tag.

**Host:** *E. coli* BL21(DE3)-R3-pRARE2

**Expression:** 10ul of BL21(DE3)-R3-pRARE2 glycerol stock were inoculated into 5ml of TB with 50ug/ml of kanamycin and 34ug/ml chloramphenicol and grown overnight at 37°C, 200rpm. 10ml of overnight culture were added to 1L of TB with 50ug/ml kanamycin and incubated at 37°C, 160rpm. After the OD<sub>600</sub> reached 1.0, the temperature was dropped to 18°C and 500ul of 1M IPTG was added to the final concentration of ~0.5mM. The culture was then incubated with shaking overnight at 18°C, 160rpm. The following morning the 4L culture was harvested and centrifuged for 10min at 4000rpm. Supernatant was discarded and cell pellets were resuspended in 80ml of a lysis buffer and frozen at -80°C.

**Extraction: Lysis buffer:** 50mM HEPES pH 7.5, 500mM NaCl, 5mM Imidazole, 5% glycerol + 1mM PMSF. The thawed cells were broken by 5 passes at 16.000 psi through a high pressure homogeniser followed by centrifugation for 45 min at 15.000rpm.

**Column 1:** Ni-affinity, His-Trap, 1 ml (Amersham)

**Column 2:** Superdex 200, HiPrep 16/60 (Amersham)

**Buffers: Binding buffer:** 50mM HEPES pH 7.5, 500mM NaCl, 20mM Imidazole, 5% glycerol, 1mM PMSF, 0.5mM TCEP; **Washing buffer:** 50mM HEPES pH 7.5, 500mM NaCl, 40mM Imidazole, 5% glycerol, 1mM PMSF, 0.5mM TCEP; **Elution buffer:** 50mM HEPES pH 7.5, 500mM NaCl, 5% glycerol, 250mM Imidazole, 0.5mM TCEP; **GF buffer:** 10mM HEPES pH 7.5, 500mM NaCl, 5% glycerol, 0.5mM TCEP

**Procedure:** The cell extract was loaded on the AKTA Express system. The extinction at 280nm was monitored and fractions were collected and analyzed by SDS-PAGE. Positive fractions were pooled for TEV cleavage.

**TEV cleavage:** The His-tag was cleaved with 1 mg TEV per 40 mg target protein at 4°C overnight. The protein was further purified on IMAC Sepharose using buffers as above.

**Concentration and buffer exchange:**

Using Amicon Ultra-15 concentrators with 30 kDa cutoff, the sample was buffer-exchanged into 10 mM Tris pH 8.5, 100 mM NaCl and concentrated to 14.91 mg/ml. Concentrations were determined from the absorbance at 280 nm using NanoDrop.

**Mass spectrometry characterization:** Calculated mass of the construct was 45105. The exact mass for the protein lacking the N-terminal methionine was confirmed by the mass spectrometry.

**Crystallization:** Crystals were grown by vapour diffusion in sitting drops at 4°C. A sitting drop consisting of 50 nl protein and 100 nl well solution was equilibrated against well solution containing 0.2M Na(acetate); 0.1M Bis-Tris Propane pH 6.5; 20% PEG 3350, 10% Ethylene Glycol. Crystals were cryo-protected in 10% Ethylene Glycol and 90% well solution before flash-cooling in liquid nitrogen.

**Data Collection: Resolution:** 1.75Å; **X-ray source:** Diamond microfocus