

PP-CDPK10

PDB:3HKO

Revision

Revision Type:created

Revised by:created

Revision Date:created

Entry Clone Accession:

Entry Clone Source:

SGC Clone Accession:cgd7_40:G126-S452:G4

Tag:N-terminal tag: mhhhhhssgrenlyfqg

Host:*E. coli* BL21(DE3)R3pACYC-LIC+LamP-phosphatase

Construct

Prelude:

Sequence:

GSLLLELQKKYHLKGAIGQGSYGVRVAIENQTRAIRAIKIMNKNKIRQINPKDVERIKTEVRLMKKLHHPNIARLYEVYEDEQYICL
VMELCHGGHLLDKLNVFIDDSTGKCAMDVVKTQICPCPECNEEAINGSIHGFRESLDFVQREKLISNIMRQIFSALHYLHNQGICHR
DIKPENFLFSTNKSFEIKLVDFGLSKEFYKLNNGEYYGMTTAKGTPYFVAPEVLNTTNESYGPKCDAWSAGVLLHLLMGAVPFP
NDADTISQVLNKKLCFENPNYNVLSPLARDLLSNLLNRNVDERFDAMRALQHPWISQFSDKIYKMS

Vector:p15-mhl

Growth

Medium:TB

Antibiotics:

Procedure:cgd7_40:G126-S452:G4 was expressed in *E. coli* BL21(DE3)R3pACYC-LIC+LamP-phosphatase cells in Terrific Broth (TB) in the presence of ampicillin/chloramphenicol (50 microg/mL and 25 microg/mL respectively). A single colony was inoculated into 10 mL of LB with of ampicillin/chloramphenicol (50 microg/mL and 25 microg/mL respectively) in a 50 mL Falcon tube and incubated with shaking at 250 rpm overnight at 37 degC. The culture was transferred into 50 mL of TB with 50 microg/mL ampicillin in a 250 mL shaking flask and incubated at 37 degC for 3 hours. Then the culture was transferred into 1.8 L of TB with 50 microg/mL kanamycin and 0.3 mL of antifoam (Sigma) in a 2 L bottle and cultured using the LEX system to an OD 600 of ~5, cooled to 15 degC, and induced with 0.5 mM isopropyl-1-thio-D-galactopyranoside (IPTG) overnight at 15 degC.

Purification

Procedure

STEP1:The cleared lysate was loaded onto a column prepacked with 10 g DE52 (Whatman)

anion exchange resin (previously activated with 2.5 M NaCl and equilibrated with Binding Buffer); and subsequently onto a 3 mL Ni-NTA (Qiagen) column pre-equilibrated with Binding Buffer at approximately 1 - 1.5 mL/min. The volume of the Ni-NTA resin was pre-determined by the predicted protein yield from test expression analysis. After the lysate was loaded, the DE52 was further washed with 20 mL of Binding Buffer. The Ni-NTA column was then washed with 200 mL of Wash Buffer at 2 - 2.5 mL/min. After washing, the protein was eluted with 15 mL of Elution Buffer. EDTA was immediately added to the elution fraction to 1 mM; and TCEP was added to 1 mM after approximately 15 more minutes.

STEP2: The sample was loaded onto a Sephadex S200 26/60 gel filtration column pre-equilibrated with 10 mM HEPES, pH 7.5 and 500 mM NaCl, 5mM DTT. The collected fractions corresponding to the correct eluted protein peak were concentrated using a 15 mL Amicon Ultra centrifugal filter device (Millipore). The protein sample identity were evaluated by mass spectroscopy. The concentrated sample (10 mg/ml) was stored at -80 degC.

Extraction

Procedure

The culture was harvested by centrifugation. Pellets from 4 L of culture were resuspended to approximately 40 mL/L of cell culture in Binding Buffer with the addition of protease inhibitors (1 mM benzamidine and 1 mM phenylmethyl sulfonyl fluoride (PMSF)). Resuspended pellets stored at -80 degC were thawed overnight at 4 degC on the day before purification. Prior to mechanical lysis, each pellet from 1 L of culture was pretreated with 0.5 % CHAPS and 500 units of benzonase for 30 minutes at room temperature. Cells were mechanically lysed with a microfluidizer (Microfluidizer Processor, M-110EH) at approximately 18000 psi; and the cell lysate was centrifuged using a Beckman JA-25.50 rotor at ~75000 x g (24000 rpms) for 20 minutes at 10 degC.

Concentration: 10 mg/ml.

Ligand

MassSpec:

Crystallization: The protein was crystallized at 20 degC in 25% w/v PEG 335, 0.2 M NaCl, 0.1M HEPES pH - 7.5, 10% Glycerol using the Hanging drop vapor diffusion method. Ligands 5mM AMPPNP, 2mM MgCl₂ added to protein before setting up plate

NMR Spectroscopy:

Data Collection:

Data Processing: