

EPHA1

PDB:3HIL

Revision

Revision Type:created

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Entry Clone Accession:NP_005223

Entry Clone Source:Open Biosystems (epha1.LIFESEQ2516475.OBS.2516475.pINCY)

SGC Clone Accession:

Tag:Nterminal tag: MHHHHHHSSGRENLYFQGD

Host:BL21 (DE3)

Construct

Prelude:

Sequence:

mhhhhhhssgrenlyfqgdGIPYRTVSEWLESIRMKRYILHFHSAGLDTMECVLELTAEDLTQMGITLPGHQKRILCSIQGF

Vector:pET28-MHL

Growth

Medium:TB

Antibiotics:

Procedure:Competent BL21 (DE3) cells (Invitrogen catalog # C6000-03) were transformed and grown using the LEX system (Harbinger BEC) at 37 degC in 2L bottles (VWR 89000-242) containing 1800 ml of TB (Sigma T0918) supplemented with 150 mM glycerol, 100 μ M Kanamycin, and 600 μ l antifoam 204 (Sigma A-8311). At OD(600) = 6, the temperature was reduced to 15 degC, and one hour later the culture was induced with 100 μ M IPTG (BioShop IPT001) and incubated overnight (16 hours) at 15 degC. Cell pellets were collected by centrifugation (12,227 xg, 20 mins) and frozen at -80 degC.

Purification

Procedure

Unclarified lysate was mixed with HisLink resin (Promega, V882A, 2.0 mL settled resin per 40 mL lysate) for 60 minutes at 4 degC. The resin was spun (500 xg for 2 minutes), batch-washed (4X45 mL of cold Wash Buffer, and transferred to a column. After additional washing (50 column volumes), protein was eluted with 40-50 mL of elution buffer and dialyzed against 50 volumes of Dialyses Buffer overnight at 4 degC. The protein sample was concentrated using a 3,000 molecular weight cut-off Amicon Ultra-15 (Millipore, UFC900524) at 4750 xg to a final concentration of 20-30 mg/mL. Protein yield was 40 mg per liter of bacterial culture. Coomassie-

stained SDS-PAGE showed that the product was pure, and Mass-spectroscopy by LCMS showed that the desired protein has the correct molecular weight.

Extraction

Procedure

After resuspension with an Ultra-Turrax T18 homogenizer (IKA Works) in 40 mL per liter bacterial culture of lysis buffer, cells were lysed by sonication (Misonix, Sonicator 3000, probe catalog # 15-338-276) on ice for 10 minutes (10 sec pulses at half-maximal frequency with 10 second rest).

Concentration: 20-30 mg/mL.

Ligand

MassSpec:

Crystallization: The crystal that was used to collect data was grown at 20 degC using the hanging drop method with equal volumes of sample and Crystallization Buffer (22% PEG3350, 0.3 M MgNO_3 . Immediately prior to setting-up crystallization plates, chymotrypsin was added to the protein sample to a final concentration of 5.7×10^{-7} M (0.57 micromolar) and protein concentration of 2.5×10^{-3} M (2.5 millimolar). Prior to dunking and storage in liquid nitrogen, suitable crystals were immersed in 1 uL CB and 1 uL cryoprotectant (20% (w/v) sucrose, 4% (w/v) glucose, 18% (v/v) glycerol and 18% (v/v) ethylene glycol).

NMR Spectroscopy:

Data Collection: Diffraction data from a crystal of the EphA1 SAM domain was collected at beamline F1 at the Cornell High Energy Synchrotron Source (CHESS) and processed using the HKL2000 program suite.

Data Processing: The structure was solved by molecular replacement techniques using the program PHASER and search model PDB entry 2QKQ. Iterative model building using the graphics program Coot, and maximum-likelihood and TLS refinement with the program REFMAC5 led to a model with an R factor of 20.8% (Rfree 27.9%) for data between 2.0-30.0 Å. Parameters for Translation/liberation/screw (TLS) refinement were generated using the TLSMD web server. The coordinates and structure factors have been deposited 2009.05.20 into the RCSB PDB database with ID code 3HIL.