

<b>Entry Clone Source:</b> MGC
<b>Entry Clone Accession:</b> IMAGE:5173213
<b>SGC Construct ID:</b> ABL2A-c055
<b>GenBank GI number:</b> gi 6382060
<b>Vector:</b> pFB-LIC-Bse. Details [ <a href="#">PDF</a> ]; Sequence [ <a href="#">FASTA</a> ] or [ <a href="#">GenBank</a> ]
<p><b>Amplified construct sequence:</b></p> <p>TACTTCCAATCCATGGACAAATGGGAAAT  GGAGCGAACAGATATTACCATGAAGCACA  AACTTGGGGGCGGTTCAGTATGGAGAGGTT  TACGTTGGCGTCTGGAAGAAATACAGCCT  TACAGTTGCTGTGAAAACATTGAAGGAAG  ATACCATGGAGGTAGAAGAATTCCTGAAA  GAAGCTGCAGTAATGAAGGAAATCAAGCA  TCCTAATCTGGTACAACCTTTTAGGTGTGT  GTACTTTGGAGCCACCATTTTACATTGTG  ACTGAATACATGCCATACGGGAATTTGCT  GGATTACCTCCGAGAATGCAACCGAGAAG  AGGTGACTGCAGTTGTGCTGCTCTACATG  GCCACTCAGATTTCTTCTGCAATGGAGTA  CTTAGAGAAGAAGAATTTTCATCCATAGAG  ATCTTGCAGCTCGTAACTGCCTAGTGGGA  GAAAACCATGTGGTAAAAGTGGCTGACTT  TGGCTTAAGTAGATTGATGACTGGAGACA  CTTATACTGCTCATGCTGGAGCCAAATTT  CCTATTAAAGTGGACAGCACCAGAGAGTCT  TGCCTACAATACCTTCTCAATTAAATCTG  ACGTCTGGGCTTTTGGGGTATTGTTGTGG  GAAATTGCTACCTATGGAATGTCACCATA  TCCAGGTATTGACCTGTCTCAGGTCTATG  ACCTACTAGAAAAAGGATATCGAATGGAA  CAGCCTGAGGGATGCCCCCTAAGGTTTA  TGAACCTATGAGAGCATGCTGGAAGTGGA  GCCCTGCCGATAGGCCCTCTTTTGCTGAA  ACACACCAAGCTTTTGAACCATGTTCCA  TGACTCTTGACAGTAAAGGTGGATA</p>
<b>Tags and additions:</b> Cleavable N-terminal His6 tag.
<p><b>Final protein sequence:</b></p> <p>mgghhhhhssgvdlgtenlyfq^SMDKWE  MERTDITMKHKLGGGQYGEVYVGWKKYS  LTVAVKTLKEDTMEVEEFLKEAAVMKEIK  HPNLVQLLGVCTLEPPFYIVTEYMPYGNL  LDYLRECNREEVTAVVLLYMATQISSAME  YLEKKNFIHRDLAARNCLVGENHVVKVAD  FGLSRLMTGDTYTAHAGAKFPIKWTAPES  LAYNTFSIKSDVWAFGVLLWEIATYGMSP  YPGIDLSQVYDLLEKGYRMEQPEGCPPKV  YELMRACWKWSPADRPSFAETHQAFETMF  HDS</p> <p>^ TEV cleave site</p>
<b>Host:</b> Baculo Virus infected Insect cells (High5 cells)

<p><b>Growth medium, induction protocol:</b>  High five cells were grown in Insect Express Medium . Cells were infected at a density of 2x10<sup>6</sup>/ml with recombinant baculovirus (virus stock P2; 1ml of virus stock/100 ml of cell culture). Cells were shaken at 120 rpm at 27°C in the innova shaker. After 48 hours post-infection the cultures were collected and centrifuged for 10min at 2000rpm. The cell pellet was resuspended in cold PBS and centrifugation was repeated. <b>Binding buffer:</b> 50 mM HEPES pH 7.5; 500 mM NaCl; 5 mM imidazole, 5% glycerol; 0.5 mM TCEP.</p>
<p><b>Extraction buffer, extraction method:</b> Frozen pellets were thawed and cells lysed using a high pressure cell disrupter. PEI (polyethyleneimine) was added to the lysate to a final concentration of 0.15 % and the lysate was centrifuged at 17,000 rpm for 30 minutes and the supernatant collected for purification.</p>
<p><b>Column 1:</b> Ni-affinity. Ni-NTA (Qiagen), 5 ml of 50% slurry in 1.5 x 10 cm column, washed with binding buffer.</p>
<p><b>Buffers:</b> <b>Binding buffer:</b> 50 mM HEPES pH 7.5, 500 mM NaCl, 5% Glycerol, 0.5 mM TCEP; <b>Wash buffer:</b> 50 mM HEPES pH 7.5, 500 mM NaCl, 20 mM Imidazole, 5% glycerol, 0.5 mM TCEP; <b>Elution buffer:</b> 50 mM HEPES pH 7.5, 500 mM NaCl, 300 mM Imidazole , 5% Glycerol, 0.5 mM TCEP.</p>
<p><b>Procedure:</b> The cleared lysate was loaded by gravity flow on the Ni-NTA column. The column was then washed with 100 ml binding buffer and 100 ml wash buffer at gravity flow. The protein was eluted by gravity flow by applying 5-ml of elution buffer.</p>
<p><b>Column 2:</b> Size Exclusion Chromatography. Superdex S200 16/60 HiLoad.</p>
<p><b>Buffers:</b> 10 mM HEPES, pH 7.5; 500 mM NaCl, , 5% Glycerol, 0.5 mM TCEP.</p>
<p><b>Procedure:</b> The protein was concentrated and applied to an S200 16/60 HiLoad gel filtration column equilibrated in 10 mM HEPES, pH 7.5; 500 mM NaCl, , 5% Glycerol, 0.5 mM TCEP using ÄKTA express system.</p>
<p><b>Mass spectrometry characterization:</b> The mass of the protein was calculated to be 33502 Da and experimentally determined mass was 33414 Da for the His tag containing protein. It is therefore likely that the difference in Mass is due to removal of the initial Met and Acetylation.</p>
<p><b>Protein concentration:</b> Protein was concentrated to 4.0 mg/ml using an Amicon 10 kDa cut-off concentrator.</p>
<p><b>Crystallization:</b> Diffraction quality crystals were grown at 4oC in 100 nl drops from a 1:1 ratio of protein and reservoir solution (20% PEG 3350; 0.1M citrate pH 5.5).</p>
<p><b>Data Collection:</b> Crystals were cryo-protected using 25% EtGly; <b>Resolution:</b> 2.05 Å; <b>X-ray source:</b> Diffraction data were collected at the SLS-X10SA.</p>