

Entry Clone Source: MGC
Entry Clone Accession: IMAGE:5199628
SGC Construct ID: MAPK11A-c007
GenBank GI number: gi 20128774
Vector: pNIC28-Bsa4. Details [PDF]; Sequence [FASTA] or [GenBank]
<p>Amplified construct sequence:</p> <p>CATATGCACCATCATCATCATCATTCTTC TGGTGTAGATCTGGGTACCGAGAACCTGT ACTTCCAATCCATGCGCGCCGGCTTCTAC CGGCAGGAGCTGAACAAGACCGTGTGGGA GGTGCCGCAGCGGCTGCAGGGGCTGCGCC CGGTGGGCTCCGGCGCCTACGGCTCCGTC TGTTCTGGCCTACGACGCCCCGGCTGCGCCA GAAGGTGGCGGTGAAGAAGCTGTCGCGCC CCTTCCAGTCGCTGATCCACGCGCGCAGA ACGTACCGGGAGCTGCGGCTGCTCAAGCA CCTGAAGCACGAGAACGTCATCGGGCTTC TGGACGTCTTCACGCCGGCCACGTCCATC GAGGACTTCAGCGAAGTGTACTTGGTGAC CACCTTGATGGGCGCCGACCTGAACAACA TCGTCAAGTGCCAGGCGCTGAGCGACGAG CACGTTCAATTCCTGGTTTACCAGCTGCT GCGCGGGCTGAAGTACATCCACTCGGCCG GGATCATCCACCGGGACCTGAAGCCCAGC AACGTGGCTGTGAACGAGGACTGTGAGCT CAGGATCCTGGATTTTCGGGCTGGCGCGCC AGGCGGACGAGGAGATGACCGGCTATGTG GCCACGCGCTGGTACCGGGCACCTGAGAT CATGCTCAACTGGATGCATTACAACCAAA CAGTGGATATCTGGTCCGTGGGCTGCATC ATGGCTGAGCTGCTCCAGGGCAAGGCCCT CTTCCCGGGAAGCGACTACATTGACCAGC TGAAGCGCATCATGGAAGTGGTGGGCACA CCCAGCCCTGAGGTTCTGGCAAAAATCTC CTCAGAACACGCCCCGACATATATCCAGT CCCTGCCCCCATGCCCCAGAAGGACCTG AGCAGCATCTTCCGTGGAGCCAACCCCT GGCCATAGACCTCCTTGGAAGGATGCTGG TGCTGGACAGTGACCAGAGGTCAGTGCA GCTGAGGCACTGGCCCACGCCTACTTCAG CCAGTACCACGACCCCGAGGATGAGCCAG AGGCCGAGCCATATGATGAGAGCGTTGAG GCCAAGGAGCGCACGCTGGAGGAGTGGA GGAGCTCACTTACCAGGAAGTCCTCAGCT TCAAGCCCTGACAGTAAAGGTGGATACGG ATCCGAA</p>
Tags and additions: Cleavable N-terminal His6 tag.
<p>Final protein sequence:</p> <p>mhhhhhssgvdlgtenlyfq^sMRAGFY RQELNKTVWEVPQRLQGLRPVGSAYGSV CSAYDARLRQKVAVKKLSRPFQSLIHARR TYRELRLLLKHLKHENVIGLLDVFTPATSI EDFSEVYLVTTLMGADLNNIVKCQALSDE</p>

<p>HVQFLVYQLLRGLKYIHSAGIIHRDLKPS NVAVNEDCELRIIDFGLARQADEEMTGYV ATRWYRAPEIMLNWMHYNQTVDIWSVGC MAELLQGKALFPGSDYIDQLKRIMEVVG PSPEVLAKISSEHARTYIQSLPPMPQKDL SSIFRGANPLAIDLLGRMLVLDSQRVSA AEALAHAYFSQYHDPEDPEAEPEYDESVE AKERTLEEWKELTYQEVLSFKP</p> <p>^ TEV cleave site</p>
Host: BL21 (DE3)R3 (Phage resistant strain)
<p>Growth medium, induction protocol: 1ml from a 10 ml overnight culture containing 50 µg/ml kanamycin was used to inoculate 1 litre of LB containing 50 µg/ml kanamycin. Cultures were grown at 37°C until the OD₆₀₀ reached ~0.3 then the temperature was adjusted to 18°C. Expression was induced for overnight using 1 mM IPTG at an OD₆₀₀ of 0.8. The cells were collected by centrifugation and the pellet resuspended in binding buffer and frozen. Binding buffer: 50 mM HEPES pH 7.5; 500 mM NaCl; 5 mM imidazole, 5% glycerol; 0.5 mM TCEP.</p>
<p>Extraction buffer, extraction method: Frozen pellets were thawed and cells lysed using a high pressure cell disrupter. PEI (polyethyleneimine) was added to the lysate to a final concentration of 0.15 % and the lysate was centrifuged at 17,000 rpm for 30 minutes and the supernatant collected for purification.</p>
<p>Column 1: Ni-affinity. Ni-NTA (Qiagen), 5 ml of 50% slurry in 1.5 x 10 cm column, washed with binding buffer.</p>
<p>Buffers : Binding buffer: 50 mM HEPES pH 7.5, 500 mM NaCl, 5% Glycerol, 0.5 mM TCEP; Wash buffer: 50 mM HEPES pH 7.5, 500 mM NaCl, 30 mM Imidazole, 5% glycerol, 0.5 mM TCEP; Elution buffer: 50 mM HEPES pH 7.5, 500 mM NaCl, 50 to 250 mM Imidazole , 5% Glycerol, 0.5 mM TCEP.</p>
<p>Procedure: The cleared lysate was loaded by gravity flow on the Ni-NTA column. The column was then washed with 100 ml binding buffer and 100 ml wash buffer at gravity flow. The protein was eluted by gravity flow by applying 5-ml portions of elution buffer with increasing concentration of imidazole (50 mM, 100 mM, 150 and 250 mM). Fractions were collected until essentially all protein was eluted, and 10 mM DTT was added.</p>
<p>Enzymatic treatment: The N-terminal His tag was cleaved by overnight incubation at 4°C with TEV protease. Cleaved products and TEV protease were removed by binding to Ni-NTA after buffer exchange to 50 mM HEPES pH 7.5, 500 mM NaCl, 5% glycerol, 0.5 mM TCEP using a 10 kDa cut-off concentrator.</p>
<p>Column 2: Size Exclusion Chromatography. Superdex S200 16/60 HiLoad.</p>
<p>Buffers: 10 mM HEPES, pH 7.5; 100 mM NaCl, 10 mM DTT</p>
<p>Procedure: The protein was concentrated and applied to an S200 16/60 HiLoad gel filtration column equilibrated in 10 mM HEPES, pH 7.5; 100 mM NaCl, 10 mM DTT using either an ÄKTA prime or ÄKTA express system.</p>
<p>Mass spectrometry characterization: LC- ESI -MS TOF gave a measured mass of 39704 for this construct as predicted from the sequence of this protein.</p>
<p>Protein concentration: Protein was concentrated to 11.8 mg/ml using an Amicon 10 kDa cut-off concentrator.</p>
<p>Crystallization: Diffraction quality crystals were grown at 4°C in 150 nl drops from a 1:1 ratio of protein and reservoir solution (20% PEG 3350; 0.1M citrate pH 5.5).</p>
<p>Data Collection: Crystals were cryo-protected using 25% EtGly; Resolution: 1.9 Å resolution limit; X-ray source: Diffraction data were collected at the Diamond beamline I04.</p>