

# CENTA1+KIF13B

PDB:3FM8

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**CENTA1: BC033747

KIF13B: BC005977

**Entry Clone Source:**CENTA1: MGC AT53-B3

KIF13B: OpenBiosystem CloneID:4103715

**SGC Clone Accession:**CENTA1: HPC060-A04

KIF13B: HPC079-B11

**Tag:**Both have N-terminal tag: mhhhhhssgrenlyfq\*g

**Host:**BL21-CodonPlus(DE3)-RIL

## Construct

**Prelude:**Tag not removed

CENTA1:M1-P374

KIF13B:G440-P545

**Sequence:**

CENTA1:

mhhhhhssgrenlyfqgMAKERRRAVLELLQRPGNARCADCGAPDPDWASYTLGVFICLSCSG  
IHRNIPQVSKVKSURLDAWEEAQVEFMASHGNDAAARAFESKVPSFYRPTPSDCQLLR  
EQWIRAKYERQEFIYPEKQEPYSAGYREGFLWKGRDNGQFLSRKFVLTEREGALKYFN  
RNDAKEPKAVMKIEHLNATFQPAKIGHPHGLQVTYLKDNSTRNIFIYHEDGKEIVDWFNA  
LRAARFHYLQVAFPGASDADLVPKLSRNYLKEGYMEKTGPKQTEGFRKRWFTMDDRRL  
MYFKDPLDAFARGEVFIGSKESGYTVLHGFPPSTQGHHPHGITIVTPDRKFLFACETESD  
QREWVAAFQKAVDRPMLPQEYAVEAHFKHKP

KIF13B:

mhhhhhssgrenlyfqgGIKVGDDKCFLVNLNADPALNELLVYYLKEHTLIGSANSQDIQLCGM  
GILPEHCIIDITSEGQVMLTPQKNTRTFVNGSSVSSPIQLHHGDRILWGNNHFFRLNLP

**Vector:**pET28-mhl (GI:134105571)

## Growth

**Medium:**Terrific Broth

**Antibiotics:**Kanamycin 50 µg/mL Chloramphenicol 25 µg/mL

**Procedure:**LEX Bubbling. The target protein was expressed in *E. Coli* by inoculating 100 mL of overnight culture grown in Luria-Bertani medium into 2L Terrific Broth medium in the presence of 50 µg/mL kanamycin and 50 µg/mL chloramphenicol at 37 degC. When OD600 reached ~3.0, the temperature of the medium was lowered to 18 degC and the culture was induced with 1 mM

IPTG. The cells were allowed to grow overnight before they were harvested and flash frozen in liquid nitrogen and stored at -80 degC.

## **Purification**

### **Procedure**

The lysate was centrifuged at 15,000 rpm for 45 minutes and the supernatants were mixed with 1.5 mL 50% Ni-NTA beads, and incubated at 4 degC for 1.5 hours. The supernatant was then passed through a gravity column (Poly-Prep, Bio-Rad, Catalog #731-1550) and the beads were washed using 10 mL washing buffer twice. The protein bound to beads were eluted using 10 mL elution buffer twice. The flow-through was collected and loaded onto Superdex-200 gel filtration column. Eluted fractions were pooled and concentrated using amicon centrifugal filter (m.w. cut-off 10,000 for CEN1A1, cut-off 5,000 for KIF13B-FHA). The purity of the proteins was higher than 95% judged by SDS-PAGE. The complex was formed by mixing purified CEN1A1 and KIF13B at 1:1 ratio and run through a Superdex-75 column. The fractions were collected and concentrated using Amicon centrifugal filter and then used to setup crystallization.

## **Extraction**

### **Procedure**

Frozen cells were thawed and resuspended in 80 mL extraction buffer with freshly added 1mM PMSF/Benzomidine, 5U/ml of Benzonase (Sigma Catalog # E1014, 250U/ $\mu$ L), 0.5% CHAPS, and supplemented with protease inhibitor cocktail (SIGMA Catalog # P8849), and lysed using microfluidizer (17000 psi).

**Concentration:** 28.2 mg/mL (complex concentration using Bradford measurement)

### **Ligand**

**ZnMassSpec:** CEN1A1: Native: 45619.63, expected 45618.66

KIF13b: Native: 13951.10, expected 13950.80

**Crystallization:** Buffer for the protein is gel filtration buffer.

Crystal used for data collection was grown in Optimized SGC-B12 condition: Well solution: 1.20 M Li<sub>2</sub>SO<sub>4</sub>, 0.1M Sodium Citrate pH 6.0 0.5M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> Protein complex was added 5% MPD and then mixed with well solution at 1uL:1uL ratio, and setup as hanging drop in a 24-well optimization plate. Crystals grown to a size of 60micron in about 3-4 weeks. Cryo used 1.6M Li<sub>2</sub>SO<sub>4</sub> and 0.5M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>.

### **NMR Spectroscopy:**

#### **Data Collection:**

#### **Data Processing:**