

# CENTA1

**PDB:**3FEH

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**BC033747

**Entry Clone Source:**MGC AT53-B3

**SGC Clone Accession:**HPC077-H02

**Tag:**N-terminal tag: mhhhhhhsgrenlyfq\*g

**Host:**BL21-CodonPlus(DE3)-RIL

## Construct

**Prelude:**Tag not removed

CENTA1:K3-F370

**Sequence:**

mhhhhhhsgrenlyfqgKERRAVLELLQRPGNARCADCAGPPDWASYTLGVFICLSCSGIHRNIPQVSKVKSVRDAWEEAQVE  
FMASHGNDARARFESKVPSFYRPTPSDCQLLREQWIRAKYERQEFIGEYREGFLWKRGRDNGQFLSRKFVLTEREG  
ALKYFNRNDAKEPKAVMKIEHLNATFQPAKIGHPHGLQVTYLKDNSTRNIFYHEDGKEIVDWFNALRAARFHQLQVAFPGASDADL  
VPKLSRNYLKEGYMEKTGPKQTEGFRKRWFTMDDRLMYFKDPLDAFARGEVFIGSKESGYTVLHGFPSTQGHWPNGITIVTPDR  
KFLFACETESDQREWVAAFQKAVDRPMLPQEYAVEAHF

**Vector:**pET28-mhl (GI:134105571)

## Growth

**Medium:**Terrific Broth

**Antibiotics:**Kanamycin 50  $\mu$ g/mL Chloramphenicol 25  $\mu$ g/mL

**Procedure:**LEX Bubbling. The target protein was expressed in *E. coli* by inoculating 100 mL of overnight culture grown in Luria-Bertani medium into 2L Terrific Broth medium in the presence of 50  $\mu$ g/mL kanamycin and 50  $\mu$ g/mL chloramphenicol at 37 degC. When OD600 reached ~3.0, the temperature of the medium was lowered to 18 degC and the culture was induced with 1 mM IPTG. The cells were allowed to grow overnight before they were harvested and flash frozen in liquid nitrogen and stored at -80 degC.

## Purification

### Procedure

The lysate was centrifuged at 15,000 rpm for 45 minutes and the supernatants were mixed with 1.5 mL 50% Ni-NTA beads, and incubated at 4 degC for 1.5 hours. The supernatant was then passed through a gravity column (Poly-Prep, Bio-Rad, Catalog #731-1550) and the beads were washed using 10 mL washing buffer twice. The protein bound to beads were eluted using 10 mL

elution buffer twice. The flow-through was collected and loaded onto Supderdex-200 gel filtration column. Eluted fractions were pooled and concentrated using amicon centrifugal filter (m.w. cut-off 10,000 ). The purity of the proteins was higher than 95% judged by SDS-PAGE.

## Extraction

### Procedure

Frozen cells were thawed and resuspended in 80 mL extraction buffer with freshly added 1mM PMSF/Benzomidine, 5U/ml of Benzonase (Sigma Catalog # E1014, 250U/ $\mu$ L), 0.5% CHAPS, and supplemented with protease inhibitor cocktail (SIGMA Catalog # P8849), and lysed using microfluidizer(17000 psi).

**Concentration:**43.3 mg/mL (14.0 mg/mL for SeMet labeled protein)

### Ligand

**ZnMassSpec:**Native: 44928.82, expected 44925.78

SeMet: 45259.89, expected 45254.01

**Crystallization:**Buffer for protein is 20mM HEPES pH7.3, 300mM NaCl, 1mM TCEP, 5% Glycerol.

Crystallization was setup using sitting drops with Red Wings and SGC-I screens initially. Plate crystals were seen in conditions RW-G08, G09 and H02. Native crystal for structure determination was grown in 25% P3350, 0.2M NH4Ac, and 0.1M Bis-Tris pH 6.5 in sitting drop setting (0.5uL protein + 0.5uL well solution), crystals usually appear in 4-7 day, cryoprotectant used 20% P3350 + 20% EG.

SeMet crystal used for structure determination was grown in 30% P3350, 0.2M NH4Ac, and 0.1M Bis-Tris pH 6.0 in hanging drop setting (1uL + 1uL). Same cryo used.

### NMR Spectroscopy:

### Data Collection:

### Data Processing: