

# TDRD2

PDB:3FDR

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**NP\_006853

**Entry Clone Source:**

**SGC Clone Accession:**

**Tag:**N-terminal hexahistidine tag with integrated TEV protease cleavage site: mhhhhhssgrenlyfq\*g. The tag was removed for crystallization.

**Host:***E. coli* BL21(DE3)-V2R-pRARE2

## Construct

**Prelude:**

**Sequence:**

gGSRSLQLDKLVNEMTQHYENSVPEDLTVHVGDIVAAPLPTNGSWYRARVLGTLENGNLDLYFVDFGDNDCPLKDLRALRSDFLSL  
PFQAI ECS

**Vector:**pET28-MHL

## Growth

**Medium:**TB

**Antibiotics:**

**Procedure:**A fresh transformation was used to inoculate 20 mL LB media containing 50 µg/mL kanamycin. The culture was grown overnight at 37°C with shaking. The next day this starter culture was used to inoculate 2L of TB medium which contained 50 µg/mL kanamycin. The culture was grown in LEX at 37°C to OD600 of 2.3 and was induced with the addition of 0.5 mM IPTG. The temperature was reduced to 16°C and the culture was incubated for a further 18 hours before harvesting the cells.

## Purification

**Procedure**

Column 1: Affinity purification, open Ni-NTA column Procedure: The supernatant was incubated with 6mL of 50% slurry Ni-NTA beads by rocking. After 1 hour incubation at 4°C, the beads were washed with 50 mL of lysis buffer. The protein was eluted using ~20mL EB.

Column 2: HiTrap Q HP 5mL Procedure: The eluent from the Ni column diluted 1:20 in Buffer A and manually loaded on the column. The protein was then eluted via a linear gradient from 0 -

100% of buffer B.

Column 3: Gel filtration, HiLoad 16/60 Superdex 75 Prep Grade Procedure: The eluent from the IEX column was loaded onto the gel filtration column in GF buffer at 1 mL/min, fraction size 2mL. The fractions containing protein were identified on a SDS-PAGE gel. Final step: The Histidine tag was removed via incubated with TEV protease and the resulting sample purified using the same conditions as column 3.

## **Extraction**

### **Procedure**

Cells were harvested by centrifugation and pellets were stored in -80°C. Prior to purification, the cell pellet was resuspended in lysis buffer. Cells were disrupted by sonication (10 minutes) and samples were centrifuged for 60 min at 70000 g.

**Concentration:** 20.25 mg/ml.

### **Ligand**

### **MassSpec:**

**Crystallization:** 30% PEG 4000, 0.2 M Ammonium Acetate, 0.1 M NaCitrate, pH 5.6

### **NMR Spectroscopy:**

### **Data Collection:**

### **Data Processing:**