

# CNDP1

**PDB:**3DLJ

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**BC117122

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**N-terminal tag: APEHHHHHHHDYDIPTTENLYFQGAMD

**Host:**Sf9 insect cells

## Construct

**Prelude:**

**Sequence:**

gamdSPSPPPALLEKVFQYIDLHQDEFVQTLKEWVAIESDSVQPVPRFRQELFRMMAVAADTLQRLGARVASVDMGPQQLPDGQSLP  
IPPVILAELGSDPTKGTVC FYGHL DVQPADRGDGLTDPYVLTEVDGKLYGRGATDNKGPVLAWINAVSAFRALEQDLPVNIKFIE  
GMEEAGSVALEELVEKEKDRFFSGVDYIVISDNLWISQRKPAITYGTRGNSYFMVEVKCRDQDFHSGTGGILHEPMADLVALLGSL  
VDSSGHILVPGIYDEVVPLTEEEINTYKAHLDLEEYRNSRVEKFLFDTKEEILMHLWRYPSSLIHGIEGAFDEPGTKTVIPGRVI  
GKFSIRLVPHMNVSAVEKQVTRHLEDVFSKRNSSNMVVSMTLGLHPWIANIDDTQYLAACKRAIRTVFGTEPDMIRDGSTIPIAKMF  
QEIVHKS SVL IPLGAVDDGEHSQNEKINRWNYIEGTLFAAFFLEMAQLH

**Vector:**pFHMSP-LIC-N

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**Plasmid transfer vector pFHMSP-LIC-N containing the gene was transformed into DH10Bac E.coli cells (Invitrogen) to obtain recombinant viral DNA. SF9 cells were transfected with Bacmid DNA using Cellfectin reagent (Invitrogen), and recombinant baculovirus was generated. Viral stock was amplified from P1 to P3.

Sf9 cells grown in HyQ® SFX Insect Serum Free Medium (Cat.# SH3027802) at density of 3 million cells per milliliter of media and with viability not less than 97 % were infected with 7 mL of P3 viral stock for each 1 L of cell culture. Cell culture medium was collected after 4 days of incubation on a shaker at 100 RPM and 27 °C when cells viability dropped to 25-45 %.

## Purification

**Procedure**

**IMAC purification:** 1.6 L of medium was mixed with 20 mL pre-equilibrated NiNTA Superflow

beads and stirred (Talboys/Troemner) for 1 hour at room temperature. The resin was transferred to a 100 mL gravity column, washed with 100 mL of Washing Buffer, and the protein was eluted with 20 mL of Elution Buffer. A second round of NiNTA batch absorption has been performed for increased protein yield.

Bound protein was eluted from the IMAC columns with Elution Buffer and loaded onto the Gelfiltration (GF) column. The chromatogram from gel filtration showed one major protein peak that consisted of CNBP1 (confirmed by SDS-PAGE analysis). The protein was then treated with TEV protease to remove the poly-histidine tag. TEV was added in the ratio of 50:1 (w/w, CNBP1:TEV). The reaction mixture was incubated at 4°C for ~2 days. Cleavage was confirmed by SDS-PAGE analysis and the TEV and tag removed by passing the sample through a 1mL HisTrap FF column which had been equilibrated with GF buffer.

## **Extraction**

### **Procedure**

The cultured medium was centrifuged at 14,000 xg for 15 minutes, and the pH of the supernatant was adjusted to 7.5 at room temperature by adding 10x Buffer\_A. Protease inhibitors were added to final concentrations of 1 mM phenylmethanesulfonyl fluoride (PMSF, Bioshop) and 2 mM benzamidine hydrochloride (Sigma).

**Concentration:** Purified protein was concentrated using 15 mL concentrators with an appropriate molecular weight cut-off (eg Amicon Ultra-15 10,000 MWCO, Millipore) to a final value of 6 mg/mL. Average yield was about 2.5 mg/L.

### **Ligand**

### **MassSpec:**

**Crystallization:** Crystallization was setup using sitting drops with Red Wings and SGC-I screens initially. Small crystals were seen at condition RW-E9. Optimization was done using hanging drop vaporization. Crystal used for structure determination were grown in: 30.0% PEG4000, 0.2M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> 0.1 Na CaCo, pH 6.5, 10mM Sr Cl<sub>2</sub>. Paratone-N was used as cryoprotectant for data-collection.

### **NMR Spectroscopy:**

### **Data Collection:**

### **Data Processing:**