

Entry Clone Source: Site directed mutagenesis

Entry Clone Accession: IMAGE:3357140

SGC Construct ID: MLYCDA-c506

GenBank GI number: gi|6912498

Vector: pNIC28-Bsa4. Details [[PDF](#)]; Sequence [[FASTA](#)] or [[GenBank](#)]

Amplified construct sequence:

CATATGCACCATCATCATCATTC
TTCTGGTGTAGATCTGGGTACCGAGA
ACCTGTACTTCCAATCCATGGACGAG
CTGCTGCGCCGCGCGGTGCCGCCGAC
GCCGGCCTACGAGCTGCGCGCAGCAA
CACCGGCGCCCGCCGAGGGTCAGTGC
GCGGACTTCGTGAGCTTCTACGGTGG
GCTGGCCGAGACGGCCAGCGGGCCG
AACTGCTGGGCCGCCTGGCGCGGGGC
TTCGGCGTGGACCACGGCCAGGTGGC
GGAGCAGAGCGCCGGCGTGCTCCATC
TGCGCCAGCAGCAGCGGGAGGCGGCG
GTGCTGCTGCAGGCCGAGGACCGGCT
GCGCTACGCGCTGGTGCCGCGCTATC
GCGGCCTCTTCCACCACATCAGCAAG
CTGGACGGCGGCGTGCGCTTCCTGGT
GCAGCTGCGGGCCGACCTGCTGGAGG
CGCAGGCCCTCAAGCTGGTGGAGGGG
CCGGACGTCCGGGAAATGAATGGGGT
GCTGAAAGGAATGCTCTCAGAATGGT
TTTCCCCCGGGTTCCTGAACCTAGAA
CGGGTTACCTGGCATTACCGTGTGA
AGTGCTTCAGAAAATCAGTGAGGCTG
AGGCTGTGCATCCTGTAAAAAACTGG
ATGGACATGAAGCGCCGCGTTGGGCC
CTACAGAAGGTGTTACTTCTTTTCTC
ACTGTTTCGACCCCTGGGGAGCCCCTG
GTCGTTTTTGACGTGGCACTGACTGG
TGACATCTCCAGCAACATCCAGGCAA
TCGTGAAGGAACATCCTCCATCAGAA
ACAGCAGCTGCTAACAAAATCACTGC
TGCGATCTTTTATTCCATCAGCTTGA
CCCAGCAGGGACTCCAAGGGGTGGAG
CTGGGAACATTCTCATAAAGCGAGT
CGTCAAGGAGTTGCAGAGAGAGTTTC
CTCACCTTGGGGTGTTTTCAAGTCTG
TCACCTATACCTGGTTTCACCAAATG
GCTTCTGGGGCTTCTGAACTCGCAA
CGAAGGAGCATGGGAGGAATGAACTC
TTTACAGATTCTGGAATGTAAGGAAAT
CTCGGAGATCACAGGTGGCCCCATTA
ACGAGACCCTCAAGCTCCTCCTCAGC
AGCAGCGAGTGGGTGCAGTCGGAGAA

GCTGGTGCGGGCGCTGCAGACTCCGC
TGATGAGGCTGTGCGCCTGGTACCTG
TATGGAGAGAAGCACCGCGGCTACGC
GCTGAACCCCGTGGCCAACTTCCACC
TGCAGAACGGGGCGGTGCTGTGGCGC
ATCAACTGGATGGCGGATGTGAGCCT
CAGAGGCATCACCGGCTCCTGCGGCC
TGATGGCCAACTACCGCTACTTCCTG
GAGGAGACGGGGCCCCAACAGCACCTC
CTACCTCGGCTCCAAGATCATCAAAG
CCTCTGAGCAGGTCCTCAGCCTAGTG
GCCCAGTTTCAAAGAAGTACAGTA
AAGGTGGATACGGATCCGAA

Final protein sequence (Tag sequence in lowercase):

mhhhhhhssgvdlgtenlyfq^smEL
LRRVPPPTPAYELRAATPAPAEGQCA
DFVSFYGGLAETAQRAELLGRLARGF
GVDHGQVAEQSAGVLHLRQQQREAAV
LLQAEDRLRYALVPRYRGLFHHISKL
DGGVRFLVQLRADLLEAQALKLVEGP
DVREMNGVLKGMLSEWFSPGFLNLER
VTWHSPCEVLQKISEAEAVHPVKNWM
DMKRRVGPYRRCYFFSHCSTPGEPLV
VLHVALTGDISSNIQAIVKEHPPSET
AAANKITAAIFYSISLTQQGLQGVEL
GTFLIKRVVKELQREFPHLGVFSSLS
PIPGFTKWLLGLLNSQTKEHGRNELF
TDSECKEISEITGGPINETLKLSS
SEWVQSEKLVRALQTPLMRLCAWYLY
GEKHRGYALNPVANFHLQNGAVLWRI
NWMADVSLRGITGSCGLMANYRYFLE
ETGPNSTSYLGSKI IKASEQVLSLVA
QFQKN

^ TEV cleavage site

Tags and additions: Cleavable N-terminal His₆-tag.

Host: BL21 (DE3)R3-pRARE2 (Phage resistant strain).

Growth medium, induction protocol: The glycerol stock of host strain BL21 (DE3)-R3-pRARE2 was used to inoculate 10ml of TB (terrific Broth) supplemented with 50µg/ml kanamycin and 35µg/ml chloramphenicol. This starter culture was grown overnight at 37°C and used to inoculate 3x 1 litre culture in TB supplemented with 100µg/ml kanamycin. The culture was grown at 37°C until the OD₆₀₀ reached ~2.0. After that the temperature was lowered to 18°C. Protein production was induced with 1 mM IPTG and recombinant MLYCD was expressed at that temperature overnight. The next day cells were harvested by centrifugation at 5000rpm for 20 minutes then the supernatant was discarded and pellets re-suspended in 70ml of 2x lysis buffer. Stored at -80°C.

Lysis buffer: 100 mM K-phosphate, pH 7.5; 1 M NaCl; 20% Glycerol; 1 mM TCEP; 1x Protease Inhibitors Cocktail Set VII (Calbiochem, 1/1000 dilution); and 15 units/ml Benzonase.

Extraction buffer, extraction method: Frozen pellets were thawed and supplemented with

TCEP, Benzonase and protease inhibitors. Cells were lysed by sonication. Nucleic acids and cell debris were removed by adding 0.15% PEI (polyethyleneimine), then centrifuged for further 30 minutes at 17,000rpm.

Column 1: Ni-affinity, HisTrap Crude FF, 5ml (GE Healthcare).

Column 1 Buffers:

Lysis buffer: 100 mM K-phosphate, pH 7.5; 1 M NaCl; 20% glycerol; 0.5 mM TCEP.

Wash buffer: 50 mM K-phosphate, pH 7.5; 500 mM NaCl; 10% glycerol; 0.5 mM TCEP; 30 mM Imidazole.

Elution buffer: 50 mM K-phosphate, pH 7.5; 500 mM NaCl; 10% glycerol; 0.5 mM TCEP; 300 mM Imidazole.

Column 1 Procedure: The clarified cell extract was loaded on the column at 4ml/min on an ÄKTA express system (GE Healthcare). The column was washed with 10 volumes of lysis buffer, 10 volumes of wash buffer, and then eluted with elution buffer at 4ml/min. The eluted peak of A₂₈₀ was automatically collected.

Column 2: Gel filtration, HiLoad 16/60 Superdex S75 prep grade, 120ml (GE Healthcare).

Column 2 Buffer: 50 mM HEPES, pH 7.5; 500 mM NaCl; 5% glycerol; 0.5 mM TCEP.

Column 2 Procedure: The eluted fraction from the Ni-affinity column was loaded on the gel filtration column equilibrated in GF buffer at 0.80ml/min. Eluted proteins were collected in 1.8ml fractions and analyzed on SDS-PAGE.

Mass spectrometry characterization:

Observed mass: 52891.5Da

Expected mass: 52891.7Da

Protein concentration: Purified protein was concentrated to 10mg/ml using a Centricon centrifugal device with a 30kDa MW cut-off, and stored in 5 mM HEPES pH 7.5, 100 mM NaCl and 1% Glycerol at -80°C.

Crystallisation: Prior to crystallization, protein was pre-incubated with 2 mM decanoyl-CoA. Crystals were grown at 20°C by sitting drop vapour diffusion. A sitting drop mixing 100nl protein (10mg/ml) and 50nl well solution (10% w/v PEG20000 and 0.1 M MES pH 6.0) was equilibrated against 20µl well solution. Crystals were cryo-protected using 25% (v/v) ethylene glycol and flash cooled in liquid nitrogen.

Data collection:

Resolution: 2.80Å.

X-ray source: Diamond Light Source beamline I02.