

Entry Clone Source: MGC

Entry Clone Accession: IMAGE:4510603

SGC Construct ID: MGC45594A-c007

GenBank GI number: gi|28557745

Vector: pNIC28-Bsa4. Details [[PDF](#)]; Sequence [[FASTA](#)] or [[GenBank](#)]

Coding DNA sequence:

```
CATATGCACCATCATCATCATCATTTCTTC
TGGTGTAGATCTGGGTACCGAGAACCTGT
ACTTCCAATCCATGCAGGGCTCCGCCATT
CCCCAAGCCATGCAGAAGCTGGTGGTGAC
CCGGCTGAGCCCCAACTTCCGCGAGGCCG
TCACCCTGAGCCGGGACTGCCCGGTGCCG
CTCCCCGGGGACGGAGACCTCCTCGTCCG
GAACCGATTTGTTGGTGTTAACGCATCTG
ACATCAACTATTTCAGCAGGCCGCTATGAC
CCCTCAGTTAAGCCTCCCTTTGACATAGG
TTTCGAAGGCATTGGGGAGGTGGTGGCCC
TAGGCCTCTCTGCTAGTGCCAGATACACA
GTTGGCCAAGCTGTGGCTTACATGGCACC
TGGTTCTTTTGCTGAGTACACAGTTGTGC
CTGCCAGCATTGCAACTCCAGTGCCCTCA
GTGAAACCCGAGTATCTTACCCTGCTGGT
AAGTGGCACCACCGCATAATCAGCCTGA
AAGAGCTCGGAGGACTGTCGGAAGGGAAA
AAAGTTTTGGTGACAGCAGCAGCTGGGGG
AACGGGCCAGTTTGCCATGCAGCTTTCAA
AGAAGGCAAAGTGCCATGTAATTGGAACC
TGCTCTTCTGATGAAAAGTCTGCTTTTCT
GAAATCTCTTGGCTGTGATCGTCCTATCA
ACTATAAACTGAACCCGTAGGTACCGTC
CTTAAGCAGGAGTACCCTGAAGGTGTCGA
TGTGGTCTATGAATCTGTTGGGGGAGCCA
TGTTTGAAGTTGGCTGTAGACGCCCTGGCT
ACGAAAGGGCGCTTGATAGTAATAGGGTT
TATCTCTGGCTACCAAACCTCCTACTGGCC
TTTCGCCTGTGAAAGCAGGAACATTGCCA
GCCAAACTGCTCAAGAAATCTGCCAGCGT
ACAGGGCTTCTTCCTGAACCATTAACCTTT
CTAAGTATCAAGCAGCCATGAGCCACTTG
CTCGAGATGTGTGTGAGCGGAGACCTGGT
TTGTGAGGTGGACCTTGGAGATCTGTCTC
CAGAGGGCAGGTTTACTGGCCTGGAGTCC
ATATTCCGTGCTGTCAATTATATGTACAT
GGGAAAAAACACTGGAAAAATTGTAGTTG
AATTACCTCACTGACAGTAAAGGTGGATA
CGGATCCGAA
```

Tags and additions: N-terminal His-tag with TEV protease cleavage site

Host: BL21(DE3)-R3-pRARE2

Expressed protein sequence (tag sequence in lowercase):

```
mhhhhhssgvdlgtenlyfqs*MQGSAI
PQAMQKLVVTRLSPNFRFAVTLSDCPVP
```

LPDGDLLVRNRFGVGNASDINYSAGRYD
PSVKPPFDIGFEGIGEVVALGLSASARYT
VGQAVAYMAPGSFAEYTVVPASIATPVPS
VKPEYLTLLVSGTTAYISLKEGLGGLSEGK
KVLVTAAAGGTGQFAMQLSKKAKCHVIGT
CSSDEKSAFLKSLGCDRPINYKTEPVGTV
LKQEYPEGVDDVYESVGGAMFDLAVDALA
TKGRLIVIGFISGYQTPTGLSPVKAGTLP
AKLLKKSASVQGFFLNHYLSKYQAAMSHL
LEMCVSGDLVCEVDLGDLSPEGRFTGLES
IFRAVNYMYMGKNTGKIVVELPH

* TEV cleave site

Growth medium, induction protocol: 10 µl of a glycerol stock was inoculated into 3ml of LB medium (supplemented with Kanamycin, 50 µg/ml) in a 15 ml culture tube and cultured at 37°C o/n in a shaking incubator (275 rpm). Next day 1 ml of o/n culture was used to inoculate 1 litre of LB medium and grown at 37°C with vigorous shaking (180 rpm) until the culture reaches an OD₆₀₀ of 1.5. Temperature was reduced to 18°C, and cells were induced with IPTG at a concentration of 1 mM, and cultivated for 16 hrs. Cells were harvested, centrifuged at 6500 rpm for 10 min, and the pellet was stored at -20°C until further use.

Extraction buffer, extraction method: Thawed cell pellets were dissolved in 30-40 ml of binding buffer (500 mM NaCl, 5% Glycerol, 50 mM HEPES pH 7.5, 5 mM Imidazole). Cells were lysed by sonication (3x 2 minutes) in a 50ml conical tube. After lysis, the cell lysate was centrifuged at 4°C for 45 minutes at 21,000 (rpm).

Column 1: Ni-NTA resin

Buffers:

Wash buffer: 500 mM NaCl, 5% Glycerol, 50 mM Tris-HCl pH 7.5, 30 mM Imidazole

Elution buffer: 500 mM NaCl, 5% Glycerol, 50 mM HEPES pH 7.5, 250mM Imidazole

Procedure: The clarified supernatant after centrifugation was passed through a Ni-NTA (2.5ml resin) column twice. The column was washed with 50 ml of wash buffer, and protein was eluted with 15 ml of elution buffer.

Column 2: Hiload 16/60 Superdex 200 prep grade 120 ml, GE Healthcare

Buffer: 10 mM HEPES, pH 7.5, 500 mM NaCl, 5 % glycerol, 0.5 mM TCEP.

Procedure: The eluted fractions from the Ni-affinity HisTrap columns were loaded on the gel filtration column in GF buffer at 1.0 ml/min. Eluted proteins were collected in 1 ml fractions

Concentration: 5 mg/ml using Vivaspin 10K concentrators

Enzymatic treatment: none

Mass spectrometry characterization: corresponds to theoretical mass, as determined by ESI-TOF MS.

Crystallization: Crystals were grown by vapour diffusion in sitting drops at 20°C. Before crystallization setup protein was incubated with 5mM of NADP and 1.5mM of Raloxifene. A sitting drop consisting of 50 nl protein and 100 nl well solution was equilibrated against well solution containing 1.6M Na-malonate pH 7.0, 1% (w/v) jeffamine-ED-2001, 0.1M Hepes pH 8.0. Crystals were mounted in the presence of 25% ethylene glycol and flash-cooled in liquid nitrogen.

Data Collection: Resolution: 1.8 Å; **X-ray source:** Diamond IO2.