

# PTK9L

**PDB:**2VAC

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**gi|6005846

**Entry Clone Source:**MGC

**SGC Clone Accession:**PTK9LA-c013

**Tag:**mhhhhhhssgvdlgtenlyfq\*s(m) TEV-cleavable (\*) N-terminal his6 tag.

**Host:**BL21(DE3)-R3-pRARE2 (A homemade phage resistant version of BL21(DE3) containing the pRARE2 plasmid from Rosetta II (DE3) cells).

## Construct

**Prelude:**

**Sequence:**

MHHHHHHSSGVDLGTENLYFQ^smGIHATEELKEFFAKARAGSVRLIKVVIEDEQQLVLGASQEPVGRWDQDYDRAVLPLDAQQPCY  
LLYRLDSQNAQGFELFLAWSPDNPSPVRLKMLYAATRATVKKEFGGGHIKDELFGTVKDDLSFAGYQKH

**Vector:**pNIC28-Bsa4.

## Growth

**Medium:**

**Antibiotics:**

**Procedure: Glycerol stock preparation:** A number of colonies from the transformation were used to inoculate 1 ml of LB media containing 50 µg/ml kanamycin and 34 µg/ml chloramphenicol, which was placed in a 37°C shaker overnight. The next day glycerol stocks were prepared from this overnight culture.

A glycerol stock was used to inoculate 10 ml of TB media containing 50 µg/ml kanamycin, which was placed in a 37°C shaker overnight. The next day this starter culture was used to inoculate 1L of TB media containing 50 µg/ml kanamycin. After growth at 37°C until the OD600 was greater than 2, the temperature was reduced to 18°C and the cells were induced by the addition of 0.5 mM IPTG. The expression was continued overnight. Two of these 1L growths were combined for the subsequent protein purification. Cells were spun and frozen at -80°C.

## Purification

**Procedure**

**Column 1:** HisTrap 5ml.

**Column 1:** 2x HiPrep 26/10 desalting columns in series.

The protein was purified using an AktaExpress system.

**Column 1 Procedure:** The clarified cell extract was passed through the column at a flow rate of 4 ml/min. The column was then washed Binding Buffer until a stable UV baseline was achieved. The column was then washed with Wash Buffer until a stable UV baseline was achieved. The protein was eluted with 25 ml of Elution Buffer.

**TEV protease digestion:** The desalting fractions containing PTK9LA were pooled and 700  $\mu$ l of TEV protease solution (about 1 mg/ml) was added. The digestion was left overnight at 4°C. After 24 hours the TEV protease digestion had not proceeded to completion so an additional 200  $\mu$ l of TEV protease solution was added and the digestion left for an additional 24 hours.

**Rebinding of impurities to Ni-NTA:** The protein was mixed with Ni-NTA resin (4 ml, pre-equilibrated into Desalting Buffer) at 4°C for 45 minutes. The resin was spun down and the supernatant filtered through a 0.45  $\mu$ M syringe filter.

## Extraction

### Procedure

The cells were thawed and resuspended in 120 ml of Lysis buffer. The resuspended cells were passed 5 times through an Emulsiflex C5 high-pressure homogeniser, collecting a final volume of approximately 170 ml after dilution with Lysis Buffer. PEI was added to a final concentration of 0.2 % and the cell debris and precipitated DNA were spun down (16000 rpm, JLA 16.250 rotor, 38400 x g, 70 min). The supernatant was filtered through a 0.45  $\mu$ M syringe filter.

**Concentration:** The TEV protease cleaved PTK9LA was concentrated to 20.8 mg/ml (measured using a nanodrop spectrometer and a calculated extinction coefficient at 280 nm), distributed into aliquots and frozen at -80°C.

### Ligand

**MassSpec:** Measured: 15163.7;

Expected: 15162.1.

**Crystallization:** Crystals grew from a 1:1 ratio of protein to precipitant solution (0.01M ZnCl<sub>2</sub>, 0.1M Tris pH 8.0, 20% PEG 6000, 10% Ethylene Glycol), using the vapour diffusion method.

### NMR Spectroscopy:

**Data Collection:** Crystals were cryo-protected by equilibration into precipitant solution containing 25% ethylene glycol, and then flash frozen in liquid nitrogen. Data was collected on a Rigaku FR-E rotating anode X-ray source at 1.7  $\text{\AA}$  resolution.

### Data Processing: