

# L3MBTL

**PDB:**2RJ2

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**NP\_056293

**Entry Clone Source:**

**SGC Clone Accession:**L3MBTL1\_3:A3-APC053

**Tag:**N-terminal hexahistidine tag with integrated TEV protease cleavage site:  
mhhhhhssgrenlyfq\*g

**Host:**E Coli BL21(DE3) RIL CodonPlus

## Construct

**Prelude:**

**Sequence:**

GEKKECWSWESYLEEQKAITAPVSLFQDSQAVTHNKNQKLGKMLEGIDPQHPSMYFILTVAEVCGYRLRLHFDGYSECHDFWVNAN  
SPDIHPAGWFEKTGHKLQPPKGYKEEFWSQYLRSTRAQAAPKHLFVSQSHSPPLGFQVGMKLEAVDRMNPSLVCVASVTDVVD  
RFLVHFDNWDDTYDYWCDPSSPYIHPVGWCQKQKPLTPPDYDPDNFCWEKYLEETGASAVPTWAFKVRPPHSFLVNMKLEAVDR  
RNPALIRVASVEDVEDHRIKIHFDDGWSHGDFWIDADHPDIHPAGWCSKTGHPLQPPLGP

**Vector:**p28a-MHL

## Growth

**Medium:**TB

**Antibiotics:**

**Procedure:**A glycerol stock was used to inoculate 20 mL LB media containing 50 µg/mL kanamycin. The culture was grown overnight at 37°C with shaking. The next day this starter culture was used to inoculate 2L of TB medium which contained 50 µg/mL kanamycin. The culture was grown in LEX at 37°C to OD600 ~1.1 and was induced with the addition of 0.5 mM IPTG. The temperature was reduced to 18°C and the culture was incubated for a further 16 hours before harvesting the cells.

## Purification

**Procedure**

**Column 1: Affinity purification, open Ni-NTA column**  
**Procedure:** The supernatant was incubated with 6mL of 50% slurry Ni-NTA beads by rocking. After 1 hour incubation at 4°C, the beads were washed with 50 mL of lysis buffer. The protein was eluted using ~20mL EB. **Column 2: Gel filtration, HiLoad 26/60 Superdex 200 Prep Grade**  
**Procedure:** The eluent from Ni column was

loaded onto the gel filtration column in GF buffer at 2.5 mL/min, fraction size 4mL. The fractions containing protein were identified on a SDS-PAGE gel.

## **Extraction**

### **Procedure**

Cells were harvested by centrifugation and pellets were stored in -80°C. Prior to purification, the cell pellet was resuspended in lysis buffer. Cells were disrupted by sonication (10 minutes twice ) and samples were centrifuged for 60 min at 70000 g. The soluble fraction was subjected to further purification by affinity and size exclusion chromatography.

**Concentration:**20mg/ml

**Ligand**

**MassSpec:**

**Crystallization:**100mM NaAc 4.6, 100mM NaAC, 5% PEG 4K

**NMR Spectroscopy:**

**Data Collection:**

**Data Processing:**