

# NAT1

PDB:2PQT

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**GI:42741671

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**N-terminal: His-tag with integrated thrombin protease site: MGSSHHHHHHSSGLVPRGS

**Host:**E.coli BL21 (DE3) codon plus RIL (Stratagene).

## Construct

**Prelude:**

**Sequence:**

```
gsgsgsDIEAYFERIGYKKS RNKLDLETLDILQH QIRAVPFENLNIHCGDAMD LGLEAIFDQVRRNRGGWCLQVNHLLYWALTTI
GFETTMLGGVYSTPAKKYSTGMIHLLQVTIDGRNYIVDAGsGRSYQM WQPLELISGKDQPQVPCVFRLTEENGFWYLDQIRREQY
IPNEEF LHSDDLLED SKYRKIYSFTLKPRTIEDFESMNTYLQTS PSSFVTSKSFCSLQTPDGVHCLVGFTLTHRRFNYKDNTDLIEFK
TLSEEEIEKVLKNIFNISLQRKLVPKHGDRFFTI
```

**Vector:**pET28a-LIC

## Growth

**Medium:**TB

**Antibiotics:**

**Procedure:**NAT1 was expressed in E.coli BL21 (DE3) codon plus RIL in Terrific Broth (TB) in the presence of 50 µg/ml of kanamycin. Cell were grown at 37oC to an OD600 of 1.5 and induced by isopropyl-1-thio-D-galactopyranoside (IPTG), final concentration 1 mM, and incubated overnight at 15oC.

## Purification

**Procedure**

The crude extract was cleared by centrifugation. The lysate was loaded onto 10 ml Chelating Sepharose column (Amersham Biosciences), charged with Ni<sup>2+</sup>. The column was washed with 10 CV of 20 mM Tris HCl buffer, pH 8.0, containing 0.5 M NaCl , 25 mM imidazole, 5% glycerol, and the protein was eluted with elution buffer (20 mM Tris HCl, pH 8.0, 0.5 M NaCl, 250 mM imidazole, 5% glycerol). Eluted protein was treated with bromoacetanilide. The protein was loaded on Superdex200 column (26x60) (Amersham Biosciences), equilibrated with 20 mM Tris HCl buffer, pH 8.0, and 0.15 M NaCl, at flow rate 4 ml/min. Thrombin (Sigma) was added to

combined fractions containing NAT1 and incubated overnight at 4°C. The protein was further purified to homogeneity by ion-exchange chromatography on Source 30Q column (10x10) (Amersham Biosciences), equilibrated with buffer containing 20 mM Tris-HCl, pH 8.5, and eluted with linear gradient of NaCl up to 500 mM concentration (20CV). Purification yield was 1.6 mg of the protein per 1L of culture.

## **Extraction**

### **Procedure**

Cells were harvested by centrifugation at 7,000 rpm. The cell pellets were frozen in liquid nitrogen and stored at -80°C. For the purification the cell paste was thawed and resuspended in lysis buffer (20 mM Tris HCl, pH 8.0, 0.5 M NaCl, 5 mM imidazol, 2 mM β-mercaptoethanol, 5% glycerol, 0.1% CHAPS) with protease inhibitor (0.1 mM phenylmethyl sulfonyl fluoride, PMSF). The cells were lysed by passing through Microfluidizer (Microfluidics Corp.) at 20,000 psi.

**Concentration:** 4.4 mg/ml

### **Ligand**

**MassSpec:** The expected mass for NAT1 is 34431.82 Da, measured mass is 34432.1363 Da.

**Crystallization:** Purified NAT1 was crystallized using the hanging drop vapor diffusion method at 20 °C by mixing 1 μl of the protein solution with 1 μl of the reservoir solution containing 30% PEG4000, 0.2 M NaOAc, 0.1 M Tris HCl, pH 8.5.

### **NMR Spectroscopy:**

#### **Data Collection:**

#### **Data Processing:**