

DUSP13

PDB:2PQ5

Revision

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Entry Clone Accession:gi|56237018

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SGC Clone Accession:DUSP13F-c010

Tag:Not- Cleavable C-terminal His 6 tag.

Host:BL21(DE3)-R3-pRARE2

Construct

Prelude:

Sequence:

MDSLQKQDLRRPKIHGAVQASPYQPPTLASLQRLLWVRQAATLNHIDEWPSLFLGDAYAARDKSCLIQLGITHVVNAAAGKFQVDT
GAKFYRGMSLEYYIEADDNPFFDLSVYFLPVARYIRAALSVPQGRVLVHCAMGVRSATLVLAFIMIYENMTLVEAIQTVQAHRI
CPNSGFLRQLQVLDNRLGRETGRFahhhh hh

Vector:pNIC-CH.

Growth

Medium:TB

Antibiotics:

Procedure:Transformed 50 μ l competent BL21(DE3)-R3-pRARE2 cells with 10 μ l of the plasmid DNA and plated out onto LB-agar plate plus 50 μ g/ml kanamycin. The next day colonies were picked out into fresh deep well blocks containing 1 ml TB + 50 μ g/ml kanamycin which were grown overnight and glycerol stocks were prepared by adding 333 ml of 60 % glycerol to 1 ml of cell suspension, which were stored at -80°C to be used for future scale up preparations. The glycerol stock was used to inoculate 10 ml of TB (Terrific Broth) supplemented with 50 μ g/ml kanamycin. This starter culture was grown overnight at 37°C and used to inoculate a 1 liter culture in the same medium. The culture was grown at 37°C until the OD600 reached ~2.0 . After that the temperature was lowered to 18°C. Protein production was induced with 1mM IPTG and recombinant DUSP13 was expressed at that temperature overnight. The next day cells were harvested by centrifugation at 4000 rpm for 15 minutes. The cell pellet was stored at -80°C.

Purification

Procedure

Column 1: Ni-affinity chromatography: HisTrap FF Crude, 1 ml (GE Healthcare).

Column 2: Size exclusion chromatography HiLoad 16/60 Superdex 200

All purification steps were carried out using an AKTAexpress system (GE Healthcare) at 7°C. The lysate was loaded on a pre-equilibrated His-trap column at 0.8 ml/min, using a standard purification method. After loading, the column was washed at 0.8 ml/min with 10 ml binding buffer, then 20 ml wash buffer, and protein was eluted with 5 ml of elution buffer. The peak fraction was collected automatically according to A280.

The DUSP13 containing fraction eluted of the Ni-affinity chromatography was automatically loaded on the SEC column at 1.2 ml/min. Eluted fractions were 95% pure as judged by SDS-PAGE.

Extraction

Procedure

The cell pellet (38 g) was re-suspended in one volume (38 ml) of 2x extraction buffer. The re-suspended cells were lysed by one passage through a Constant Systems cell breaker and subsequent sonication; the cell breaker was washed with 1x extraction buffer, bringing the total volume to 120 ml. DNA was precipitation by addition of PEI (pH 7.5) to a final concentration of 0.15 % during an incubation time of 30 min on ice, followed by a centrifugation at 17,000 rpm (4°C); The supernatant was further cleared by filtration through a 0.2 µm serum Acrodisc filter. **Concentration:** 7 mg/ml in SEC buffer using a c entricon with a 10kDa cut off.

Ligand

MassSpec: ESI-MS revealed that the protein had the expected mass 23103 mass units.

Crystallization: DUSP13 was crystallized at 20°C using the sitting-drop vapor diffusion method. Diffraction quality crystals were obtained by mixing 7 mg/ml of protein solution with 25% PEG3350, 0.15 M ammonium citrate pH 5.0 in a 1:1 ratio.

NMR Spectroscopy:

Data Collection: Crystals were soaked in 25% Ethylene glycol + crystallization buffer before they were flash frozen in liquid nitrogen. Diffraction data were collected to 2.3 Å at the Swiss light source beam-line X10SA at a single wavelength of 1.03310 Å.

Data Processing: