

# HAT1

**PDB:**2P0W

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**GI:4504341

**Entry Clone Source:**MGC

**SGC Clone Accession:**HAT1\_05:APC038-C3

**Tag:**N-terminal: His-tag with integrated thrombin protease site: MGSSHHHHHHSSGLVPR\*GS

**Host:** E.coli BL21 (DE3) codon plus RIL (Stratagene)

## Construct

**Prelude:**

**Sequence:**

gsKKLAEYKCNTNTAIELKLVRFPELNDIRTFPEYTHQLFGDDETAFGYKGLKILLYYIAGSLSTMFRVEYASKVDENFDCVEA  
DDVEGKIRQIIPPGFCTNTNDFLSLLEKEVDFKPFGTLLHTYSVLSPTGGENFTFQIYKADMTCRGFREYHERLQTFLMWFJETASF  
IDVDDERWHYFLVFEKYNKDGLTATVGYMTVYNYVYPDKTRPRVSQMLILTPFQGGHGAQLLETVHRYYTEFPTVLDITAEDP  
SKSYVKLRDFVLVKLCQDLPCFSREKLMQGFNEDMAIEAQQKFINKQHARRVYEILRLLVTD

**Vector:**p28a-LIC

## Growth

**Medium:**TB

**Antibiotics:**

**Procedure:**HAT1 was expressed in E.coli BL21 (DE3) codon plus RIL in 6L Terrific Broth (TB) in the presence of 50 microG/ml of kanamycin. Cell were grown at 37°C to an OD600 of 1.5 and induced by isopropyl-1-thio-D-galactopyranoside (IPTG), final concentration 1 mM, and incubated overnight at 15degC.

## Purification

**Procedure**

The crude extract was cleared by centrifugation at ~75000 x g for 20 minutes. The clarified lysate was loaded onto 5 ml HiTrap Chelating column (Amersham Biosciences), charged with Ni<sup>2+</sup>. The column was washed with 10 CV of 20 mM Tris-HCl buffer, pH 8.0, containing 250 mM NaCl and 50 mM imidazole, 5% glycerol, and the protein was eluted with elution buffer (20 mM Tris-HCl, pH 8.0, 250 mM NaCl, 250 mM imidazole, 5% glycerol). The protein was loaded on Superdex200 column (26x60) (Amersham Biosciences), equilibrated with 20 mM Tris-HCl buffer, pH 8.0, and 150 mM NaCl, at flow rate 4 ml/min. Thrombin (Sigma) was added to

combined fractions containing HAT1 and incubated overnight at 4°C. The protein was further purified to homogeneity by ion-exchange chromatography on Source 30Q column (10x10) (Amersham Biosciences), equilibrated with buffer 20 mM Tris-HCl, pH 8.0, and eluted with linear gradient of NaCl up to 500 mM concentration (20CV). Purification yield was 3.8 mg of the protein per 1L of culture.

## **Extraction**

### **Procedure**

Cells were harvested by centrifugation. The cell pellets were frozen in liquid nitrogen and stored at -80°C. For the purification, 6L of cells were thawed and resuspended in lysis buffer with protease inhibitor (0.1 mM phenylmethyl sulfonyl fluoride, PMSF). The cells were lysed by passing through a Microfluidizer (Microfluidics Corp.) at 20,000 psi.

**Concentration:** 23.3 mg/ml

### **Ligand**

**MassSpec:** Expected mass = 37924.22 Da, measured mass = 38095.6689 Da.

**Crystallization:** Purified HAT1 was complexed with AcCoA (Sigma) and histone H4 peptide (SGRGKGGKGLGKGGAKRHRK) at 1:10 molar ratio of protein: AcCoA/H4 and crystallized using the sitting drop vapor diffusion method by mixing 1 ul of protein solution with 1 ul of the reservoir solution containing 12% PEG 20K, 0.1M MES pH6.5.

### **NMR Spectroscopy:**

#### **Data Collection:**

#### **Data Processing:**