

<b>Entry Clone Source:</b> TKC
<b>Entry Clone Accession:</b> n/a
<b>SGC Construct ID:</b> AK1A-c001
<b>GenBank GI number:</b> gi 4502011
<b>Final protein sequence:</b>  mhhhhhhhssgvdlgtenlyfqSMEEKLKKTNIIFV VGPGSGKGTQCEKIVQKYGYTHLSTGDLRSEVS SGSARGKKLSEIMEKGQLVPLETVLDMLRDAMVAK VNTSKGFLIDGYPREVQQGEEFERRIGQPTLLLYV DAGPETMTQRLKRGETSGRVDDNEETIKKRLETY YKATEPVIAFYEKRGIVRKVNAEGSVDSVFSQVCT HLDALLN
<b>Vector:</b> pLIC- SGC1. Details [ <a href="#">PDF</a> ]; Sequence [ <a href="#">FASTA</a> ] or [ <a href="#">GenBank</a> ]
<b>Tags and additions:</b> mhhhhhhhssgvdlgtenlyfq*s (m) TEV-cleavable (*) N-terminal his6 tag.
<b>Host:</b> BL21 (DE3)
<b>Growth medium, induction protocol:</b> Grow starter cultures from freshly transformed colonies in 10 ml LB, 0.1 mg/ml amp. This started culture was diluted 1:1000 in fresh media and was grown at 37°C to a OD <sub>600</sub> of 0.3 and than transferred to 18°C. Expression was induced at an OD <sub>600</sub> of 0.6 using 1 mM IPTG. Cells were harvested after 4h by centrifugation, transferred to 50-ml tubes, and frozen in -20oC.
<b>Extraction buffer, extraction method:</b> Extraction buffer: 50 mM Tris-HCl, pH 7.5, 500 mM NaCl. The cell pellets (4 gr wet wt) were re-suspended in 50 ml extraction buffer containing a Protease Inhibitor Coctail tablet (Roche), and lysed by a high pressure cell disrupter. Supernatant was centrifuged for 30 minutes at 17 rpm in a JA 25.5 rotor
<b>Column 1 :</b> DE52/Ni-NTA
<b>Buffers:</b> <b>Loading buffer:</b> 50 mM Hepes, pH 7.5, 500 mM NaCl, 10 mM imidazole, 5% glycerol. <b>Wash buffer:</b> 50 mM Hepes, pH 7.5, 500 mM NaCl, 20 mM imidazole, 5% glycerol. <b>Elution buffer:</b> 50 mM Hepesl, pH 7.5, 500 mM NaCl, 50-250 mM imidazole, 5% glycerol.
<b>Procedure:</b> Gravity feed chromatography. Sample applied to a 10 ml DE-52 column and washed through with 50 ml loading buffer. The flow through was applied to a 1 ml Ni-NTA column, the Ni-NTA column was washed with 2x10ml of wash buffer and eluted with elution buffer in 5 ml aliquots (Step elution using 50, 100, 150, 200 and 250 mM imidazole in the Elution Buffer)
<b>Enzymatic treatment:</b> Treated the IMAC elution(s) with TEV protease overnight.
<b>Column 2:</b> SEC
<b>Procedure :</b> AKTA-prime. Fractions containing AK1A collected from IMAC and treated with TEV protease overnight were concentrated to 1.5ml and directly applied to a S75 16/60 column equilibrated in 10 mM Hepes pH 7.5, 100 m NaCl. Flow rate 1ml/min.
<b>Mass spec characterization:</b> LC- ESI -MStof confirmed the correct mass expected for this construct.
<b>Protein concentration:</b> Centricons 10 kDa cut off
<b>Crystallization: 1Z83:</b> Crystals were grown at 4°C in 500nl sitting drops mixing 250 nl of AK1 (14 mg/ml in 10mM Hepes pH 7.5, 100mM NaCl, 10mM DTT) with 250 nl of a solution containing 20% PEG 550, 0.005 M ZnSO <sub>4</sub> , MES pH 6.5 and 1mM AP 5 A (Diadenosine pentaphosphate pentalithium salt - CAS # 94108-02-8). <b>2C95:</b> Crystal were obtained at 4 °C in sitting drops by mixing 100 nl AK1 [20

mg/ml, in 10mM HEPES pH=7.5, 100mM NaCl, 10mM DTT, 1 mM ZnSO<sub>4</sub>, 1 mM AP4A (diadenosine tetraphosphate) ] with 50 nl of the reservoir solution 2.4 M Na-malonate pH=7.0.

**Data Collection: 1Z83: Resolution: 1.9Å, X-ray source: SLS X06SA, single wavelength. 2C95: Resolution: 1.7Å, X-ray source: rotating anode (Rigaku FR-E SuperBright), single wavelength.**