

# PAK6

PDB:2C30

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**gi|9910476

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**Tag sequence: mhhhhhssgvdlgtenlyfq\*s(m) TEV-cleavable (\*) N-terminal his6 tag.

**Host:**BL21 (DE3)

## Construct

**Prelude:**

**Sequence:**

MQDPTVAKGALAGEDTGVTHEQFKAALRMVVDQGDPRLLLD SYVKIGEGSTGIVCLAREKHSGRQVAVKMMDLRKQQRRELLFNEV  
VIMRDYQHFNVVEMYKSYLVGEELWVLMFLQGGALTDIVSQVRLNEEQIATVCEAVLQALAYLHAQGV IHRDIKSDSILLTLDGRV  
KLSDFGFCQAQISKDVPKRKSLVGTPYWMAPEVISRSLYATEVDIWSLGIMVIEMVDGEPPYFSDSPVQAMKRLRDSPPPKLKNSHKV  
SPVLRDFLERMLVRDPQERATAQEELDHPFLLQTGLPECLVPLIQLYRKQTST

**Vector:**pLIC-SGC1

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**1ml from a 10 ml overnight culture containing 50 µg/ml kanamycin was used to inoculate 1 litre of LB containing 50 µg/ml kanamycin. Cultures were grown at 37°C until the OD600 reached ~0.3 then the temperature was adjusted to 18°C. Expression was induced for 4 hours using 1 mM IPTG at an OD600 of 0.8. The cells were collected by centrifugation and the pellet resuspended in binding buffer and frozen. Binding buffer: 50mM HEPES pH 7.5; 500 mM NaCl; 5 mM imidazole, 5% glycerol.

## Purification

**Procedure**

Column 1: Ni-affinity. Ni-NTA (Qiagen), 5 ml of 50% slurry in 1.5 x 10 cm column, washed with binding buffer.

## Extraction

**Procedure**

Frozen pellets were thawed and cells lysed using a high pressure cell disrupter. The lysate was centrifuged at 50,000 rpm for 40 minutes and the supernatant collected for purification.

**Concentration:****Ligand****MassSpec:**

**Crystallization:** Protein concentration, 10 mg/ml. Well solution: 1.60M magnesium sulfate, 0.1M MES 6.5.

**NMR Spectroscopy:****Data Collection:****Data Processing:**