

# NSUN5

**PDB:**2B9E

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**GI:8922322

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**N-terminal: His-tag with integrated thrombin protease site: MGSSHHHHHSSGLVPRGS

**Host:**E.coli BL21 (DE3) codon plus RIL (Stratagen).

## Construct

**Prelude:**

**Sequence:**

gsRPGPASQLPRFVRVNTLKTCSDDVVYFKRQGFSYQGRASSLDDL RALKGKHLLDPLMPELLVFPAQTDLHEHPLYRAGHLILQ  
DRASCLPAMLLDPPPGSHVIDACAAPGNKTSHAAALLKNQGKIFAFDLDKRLASMATLLARAGVSCELAEEDFLAVSPSDPRYHE  
VHYILLDPSCSGSGMPSRQLEEPGAGTPSPVRLHALAGFQQRALCHALTFFPSLQRLVYSTCSLCQEENEDVVRDALQQNPGAFRLAP  
ALPAWPHRGLSTFPGAEHCLRASPETTLSSGFFVAVIEREVPRARG

**Vector:**p28a-LIC

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**NSUN5 was expressed in E.coli BL21 (DE3) codon plus RIL in Terrific Broth (TB) in the presence of 50 µg/mL of kanamycin. Cell were grown at 37oC to an OD600 of 1.5 and induced by isopropyl-1-thio-D-galactopyranoside (IPTG), final concentration 1 mM and incubated overnight at 15oC.

## Purification

**Procedure**

The crude extract was cleared by centrifugation and passing through 20-mL DE52 column equilibrated in 20 mM HEPES, pH 7.4, containing 500 mM NaCl and 5% glycerol. The lysate was loaded onto 5 mL HiTrap Chelating column (Amersham Biosciences), charged with Ni2+. The column was washed with 10 CV of 20 mM HEPES buffer, pH 7.4, containing 500 mM NaCl , 50 mM imidazole and 5% glycerol, and the protein was eluted with elution buffer (20 mM HEPES, pH 7.4, 500 mM NaCl, 250 mM imidazole, 5% glycerol). The protein was loaded on Superdex200 column (26x60) (Amersham Biosciences), equilibrated with 20 mM Tris-HCl

buffer, pH 8.0, and 150 mM NaCl, at flow rate 4 mL/min. Thrombin (Sigma) was added to combined fractions containing NSUN5 and incubated overnight at 4°C. The protein was further purified to homogeneity by ion-exchange chromatography on Source 30Q column (10x10) (Amersham Biosciences), equilibrated with buffer 20 mM Tris-HCl, pH 8.0, and eluted with linear gradient of NaCl up to 500 mM concentration (20CV). Purification yield was 8.3 mg of the protein per 1L of culture.

## **Extraction**

### **Procedure**

Cells were harvested by centrifugation at 7,000 rpm. The cell pellets were frozen in liquid nitrogen and stored at -80°C. For the purification the cell paste was thawed and resuspended in lysis buffer (50 mM HEPES, pH 7.4, 500 mM NaCl, 5 mM imidazol, 2 mM  $\beta$ -mercaptoethanol, 5% glycerol) with protease inhibitor (0.1 mM phenylmethyl sulfonyl fluoride, PMSF). The cells were lysed by passing through Microfluidizer (Microfluidics Corp.) at 20,000 psi.

### **Concentration:**

### **Ligand**

### **MassSpec:**

**Crystallization:** Purified NSUN5 was complexed with S-adenosyl-L-methionine (SAM) (Sigma) at 1:10 molar ratio of protein:SAM and crystallized using the sitting drop vapor diffusion method at 20 °C by mixing 1  $\mu$ l of the protein solution with 1  $\mu$ l of the reservoir solution containing 10% Iso-propanole, 20% PEG 4000, 0.1M Na HEPES pH 7.5.

### **NMR Spectroscopy:**

### **Data Collection:**

### **Data Processing:**