

# ARF5

**PDB:2B6H**

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**NP\_001653

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**N-terminal: His-tag with integrated thrombin protease site before the last Ser:

MGSSHHHHHSSGLVPRGS

**Host:**E. coli BL21 (DE3)

## Construct

**Prelude:**

**Sequence:**

mgsshhhhhsglvprgsLFSRIFGKKQMRILMGLDAAGKTTIYKLKLGEIVTTIPTIGFNVEYKNICFTVWDVGGQDKIR  
PLWRHYFQNTQGLIFVVDSNDRERVQESADELQKMLQEDELRAVLLFANKQDMPNAMPVSELTDKGLQHLSRTWYVQATCATQ  
GTGLYDGLDWLSHELSKR

**Vector:**p28a-LIC

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**We prepared the seeds by inoculating freshly transforming E. coli cells (BL21 DE3) into 80 mL of Luria-Bertani medium. After overnight growth, all of the seeds were inoculated into 1.8 L of Terrific Broth medium in the presence of 50  $\mu$ g/mL of kanamycin at 37°C and grown to an OD600 of 2.28. Cells were then induced by isopropyl-1-thio-D-galactopyranoside at the final concentration of 1.5 mM and grown overnight at 20°C in the SGC LEX bubbling system.

## Purification

**Procedure**

The supernatant was passed through DE52 (Whatman) column equilibrated with the binding buffer and then loaded onto 3 mL Ni-NTA column (Qiagen) equilibrated with the same binding buffer at 4°C. The Ni-NTA column was washed with 150 mL of the wash buffer (10mM Tris pH7.5, 0.5 M NaCl, 5% glycerol, 30 mM imidazole) and the protein was eluted with 15 mL of the elution buffer (10mM Tris pH7.5, 0.5 M NaCl, 5% glycerol, 250 mM imidazole). The eluate was dialyzed overnight against a buffer containing 10 mM Tris pH 7.5, 0.5 M NaCl, 5% glycerol.

The protein concentration was estimated based on Bradford assay. Five molar equivalents of GDP, 10 mM DTT and 5 mM MgCl<sub>2</sub> were added to the purified protein before concentration. The protein was concentrated using an Amicon Ultra centrifugal filter to the final volume of 1 mL and the concentration of 20.0 mg/mL. About 20 mg of protein was obtained from 1.8 L of cell culture.

## **Extraction**

### **Procedure**

Cultures were centrifuged and the cell pellets were suspended in 100 mL of the binding buffer (10 mM Tris pH 7.5, 0.5 M NaCl, 5% glycerol, 5 mM imidazole) with a protease inhibitor cocktail (0.1 mM M benzamidine-HCl and 0.1 mM phenylmethyl sulfonyl fluoride) and flash frozen. The thawed cell pellet was lysed by a combination of 0.5% CHAPS (Sigma) and sonication. The lysate was centrifuged at 15000 rpm for 30 min and the supernatant was used for subsequent steps of purification.

### **Concentration:**

### **Ligand**

### **MassSpec:**

**Crystallization:** Purified ARF5 was crystallized using the sitting drop vapor diffusion method at room temperature. Crystals grew in one day when the protein (20 mg/mL) was mixed with the reservoir solution in a 1:1 volume ratio, and the drop was equilibrated against a reservoir solution containing 25% PEG 4K, 0.2 M AmSO<sub>4</sub>, 0.1 M Tris pH 8.5, 3% w/v 6-aminocaproic acid at 4°C. The crystals were flash frozen with the mother liquor with 15% glycerol.

### **NMR Spectroscopy:**

### **Data Collection:**

### **Data Processing:**