

# UXS1

**PDB:**2B69

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**UXS1A-s001

**Entry Clone Source:**synthetic, codon optimized for E.coli expression

**SGC Clone Accession:**

**Tag:**

**Host:**B834(DE3) (Methionine auxotroph)

## Construct

**Prelude:**

**Sequence:**

mgsshhhhhssgrenlyfqghmEKDRKRILITGGAGFVGSHLTDKLMMDGHEVTVDNFFTGRKRNV EHWIGHENFELINHDVVEP  
LYIEVDQIYHLASPPNNYMYNPIKTLKTNLTIGTLNMLGLAKRVGARLLLASTSEVYGDPEVHPQSE  
DYWGHVNPIGPRACYDEGK  
RVAETMCYAYMKQEGVEVRVARIFNTFGPRMHMNDGRVVSNFILQALQGEPLTVYSGSQTRAFQYVSDLVNGLVALMNSNVSSPVN  
LGNPEEHTILEFAQLIKNLVSGSGEIQFSEAQDDPQKRKPDIKKAKLMLGWEPVVPLEEGLNKAIHYFRKELEYQANNQgs

**Vector:**p11

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**B834(DE3) cells were transformed with the UXS1A plasmid and grown on Lb Amp plates. A single colony was used to start a 5mL overnight culture in LB/Amp, and 1mL of the overnight culture was used to inoculate 50mL of LB medium, grown at 37°C to an OD of 1.0. The cells were harvested by centrifugation, washed four times in MD medium (SelenoMet Medium Base, SelenoMet Nutrient, selenomethionine to final concentration of 40 mg/L, Molecular Dimensions) and finally the cells were inoculated in 1L of MD medium, and grown at 37°C to an OD 0.6. The culture was induced by the addition of IPTG to a final concentration of 1mM, cultured overnight at 25°C, and collected by centrifugation.

## Purification

**Procedure**

Column 1 : Ni-NTA resin

Buffers (adjusted to pH 8.0): Lysis buffer: 5 mM Imidazole, 500mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol; Wash buffer: 30 mM Imidazole, 500 mM NaCl, 50mM HEPES, pH 7.5, 5%

glycerol; Elution Buffer: 250 mM Imidazole, 500 mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol.

Column 2: Superdex S200

Buffer: 10 mM HEPES, pH 7.5, 500 mM NaCl, 5% glycerol, 1mM TCEP

Procedure: Sample was loaded, washed with wash buffer and eluted in elution buffer. The protein was concentrated using an Amicon Ultra device.

## **Extraction**

### **Procedure**

Pellets were resuspended in 20 mL lysis buffer including Protease inhibitor (complete, Roche), lysed by French Press, and the solution was centrifuged to obtain a clear supernatant (30 min, 20.000 x g). Supernatants were processed in a 2 step chromatographic procedure.

### **Concentration:**

### **Ligand**

### **MassSpec:**

**Crystallization:** Column 1 : Ni-NTA resin

Buffers (adjusted to pH 8.0): Lysis buffer: 5 mM Imidazole, 500mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol; Wash buffer: 30 mM Imidazole, 500 mM NaCl, 50mM HEPES, pH 7.5, 5% glycerol; Elution Buffer: 250 mM Imidazole, 500 mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol.

Column 2: Superdex S200

Buffer: 10 mM HEPES, pH 7.5, 500 mM NaCl, 5% glycerol, 1mM TCEP

Procedure: Sample was loaded, washed with wash buffer and eluted in elution buffer. The protein was concentrated using an Amicon Ultra device.

### **NMR Spectroscopy:**

### **Data Collection:**

### **Data Processing:**