

UXS1

PDB:2B69

Revision

Revision Type:created

Revised by:created

Revision Date:created

Entry Clone Accession:UXS1A-s001

Entry Clone Source:synthetic, codon optimized for E.coli expression

SGC Clone Accession:

Tag:

Host:B834(DE3) (Methionine auxotroph)

Construct

Prelude:

Sequence:

```
mgsshhhhhhssgrenlyfqghmEKDRKRILITGGAGFGSHLTDKLMMDGHEVTVDNFFTGRKRNVHIGHENFELINHDVVEP  
LYIEVDQIYHLASPASPPNYMYNPIKTLKTNTIGTLNMLGLAKRKGARLLLASTSEVYGDPEVHPQSEDYWGHNPIGPRACYDEGK  
RVAETMCYAYMKQEGVEVRVARIFNTFGPRMHMNDGRVVSNFIQLALQGEPLTVYGSQTRAFQYVSDLVNGLVALMNSNVSPVN  
LGNPEEHTILEFAQLIKNLVSGSEIQFSEAQDDPQKRKDIIKKAKLMLGWEPPVPLEEGLNKAIHYFRKELEYQANNQgs
```

Vector:p11

Growth

Medium:

Antibiotics:

Procedure:B834(DE3) cells were transformed with the UXS1A plasmid and grown on Lb Amp plates. A single colony was used to start a 5mL overnight culture in LB/Amp, and 1mL of the overnight culture was used to inoculate 50mL of LB medium, grown at 37°C to an OD of 1.0. The cells were harvested by centrifugation, washed four times in MD medium (SelenoMet Medium Base, SelenoMet Nutrient, selenomethionine to final concentration of 40 mg/L, Molecular Dimensions) and finally the cells were inoculated in 1L of MD medium, and grown at 37°C to an OD 0.6. The culture was induced by the addition of IPTG to a final concentration of 1mM, cultured overnight at 25°C, and collected by centrifugation.

Purification

Procedure

Column 1 : Ni-NTA resin

Buffers (adjusted to pH 8.0): Lysis buffer: 5 mM Imidazole, 500mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol; Wash buffer: 30 mM Imidazole, 500 mM NaCl, 50mM HEPES, pH 7.5, 5%

glycerol; Elution Buffer: 250 mM Imidazole, 500 mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol.

Column 2: Superdex S200

Buffer: 10 mM HEPES, pH 7.5, 500 mM NaCl, 5% glycerol, 1mM TCEP

Procedure: Sample was loaded, washed with wash buffer and eluted in elution buffer. The protein was concentrated using an Amicon Ultra device.

Extraction

Procedure

Pellets were resuspended in 20 mL lysis buffer including Protease inhibitor (complete, Roche), lysed by French Press, and the solution was centrifuged to obtain a clear supernatant (30 min, 20.000 x g). Supernatants were processed in a 2 step chromatographic procedure.

Concentration:

Ligand

MassSpec:

Crystallization: Column 1 : Ni-NTA resin

Buffers (adjusted to pH 8.0): Lysis buffer: 5 mM Imidazole, 500mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol; Wash buffer: 30 mM Imidazole, 500 mM NaCl, 50mM HEPES, pH 7.5, 5% glycerol; Elution Buffer: 250 mM Imidazole, 500 mM NaCl, 50 mM HEPES, pH 7.5, 5% glycerol.

Column 2: Superdex S200

Buffer: 10 mM HEPES, pH 7.5, 500 mM NaCl, 5% glycerol, 1mM TCEP

Procedure: Sample was loaded, washed with wash buffer and eluted in elution buffer. The protein was concentrated using an Amicon Ultra device.

NMR Spectroscopy:

Data Collection:

Data Processing: