

# ME1

**PDB:**2AW5

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**ME1A-s001

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**N-terminal His tag with TEV cleavage site

**Host:**E.coli BL21DE3 R3

## Construct

**Prelude:**

**Sequence:**

mhahhhhhsgvdlgtenlyfqsmQRGYLLTRNPHLNKDLAFTLEERQQLNIHGLLPPSFNSQEIQVLRVVKNFEHNSDFDRYLLLM  
DLQDRNEKLFYRVLTS DIEKFMPIVYPTVGLACQQYSLVFRKPRGLFITIHDRGHIASVLNAWPEDVIKAIIVTDGERILGLGDLG  
CNGMGIPVGKLALYTACGGMNPQECLPVI LDVGTENEELLKDPLYIGLRQRRVGSEYDDFLDEFMEAVSSKYGMNCLIQFEDFANV  
NAFRLLNKYRNQYCTFNDI QGTASVAVAGLLAALRITKNKLSDQTILFQGAGEAALGIAH LIVMALEKEGLPKEAIKKIWLVD SK  
GLIVKG RASLTQEKEKFAHEHEEMKNLEAIVQEIKPTALIGVAIIGGAFSEQILKDMAAFNERPIIFALSNPTS KAECSAEQCYKIT  
KGRAIFASGSPFDPVTLPNGQTLYPGQGNNSYVFPGVALGVVACGLRQITDNIFLTAEVIAQQVSDKHLEEGRLYPP LNTIRDVSL  
KIAEKIVKDAYQEKTATVYPEPQNKEAFVRSQMYSTDYDQILPDCYSWPEEVQ

**Vector:**pNIC-BSA4

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**Cells were grown for 14 hrs at 37°C in 10 mL TB medium, supplemented with Kanamycin (50 µg/mL), and were inoculated into 1 L TB medium with the same antibiotic supplement. The culture was grown at 37°C with shaking at 250 rpm. At an OD of 0.5 cells were induced with 1 mM IPTG and the temperature was lowered to 25°C. Growth was continued for an additional 18 hrs until a final OD of 8-10. Cells were collected by centrifugation, and the cell pellet was stored at -20°C until further use.

## Purification

**Procedure**

Column 1: 1 mL HisTrap FF.

Buffers: Affinity binding buffer: 10mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO4, 0.5mM TCEP; Affinity wash buffer: 50mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO4, 0.5mM TCEP; Affinity Elution Buffer: 250mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO4, 0.5mM TCEP.

Procedure: The supernatant from the last extraction step was loaded on a Akta Express system, consisting of IMAC and gel filtration as consecutive chromatography steps. Before sample loading, columns were equilibrated with the buffers indicated (HisTrap with Affinity binding buffer; gel filtration with Gel Filtration Buffer). Fractions were collected and analysed by mass spectrometry and SDS / PAGE . Peaks after gel filtration were collected, pooled and concentrated using Centricon membrane (30 kDa MW cutoff) concentrators.

Column 2 : Superdex S75

Buffers : Gel Filtration Buffer: 10mM pH7.4 Hepes, 500mM NaCl, 5% glycerol, 0.5mM TCEP.

## Extraction

### Procedure

Pellets were resuspended in approximately 3x volume of lysis buffer, and thawed at 37°C in a water bath. Cells were mechanically disrupted in a French Press, and further sonicated in 10 sec intervals for 2 mins. PEI was added to a final concentration of 0.15%, and DNA was precipitated on ice for 30 mins. The suspension was centrifuged for 17.000 rpm for 30 mins at 4°C, and the supernatant was filtered through a 0.2 um Serum Scrodisc filter. Lysis buffer: 10mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO4 , 0.5mM TCEP, 1x complete PI EDTA free tablet/50mL.

### Concentration:

#### Ligand

#### MassSpec:

**Crystallization:**Column 1: 1 mL HisTrap FF.

Buffers: Affinity binding buffer: 10mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO4, 0.5mM TCEP; Affinity wash buffer: 50mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO4, 0.5mM TCEP; Affinity Elution Buffer: 250mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO4, 0.5mM TCEP.

Procedure: The supernatant from the last extraction step was loaded on a Akta Express system, consisting of IMAC and gel filtration as consecutive chromatography steps. Before sample loading, columns were equilibrated with the buffers indicated (HisTrap with Affinity binding buffer; gel filtration with Gel Filtration Buffer). Fractions were collected and analysed by mass spectrometry and SDS / PAGE . Peaks after gel filtration were collected, pooled and concentrated using Centricon membrane (30 kDa MW cutoff) concentrators.

Column 2 : Superdex S75

Buffers : Gel Filtration Buffer: 10mM pH7.4 Hepes, 500mM NaCl, 5% glycerol, 0.5mM TCEP.

### NMR Spectroscopy:

### Data Collection:

### Data Processing: