

ME1

PDB:2AW5

Revision

Revision Type:created

Revised by:created

Revision Date:created

Entry Clone Accession:ME1A-s001

Entry Clone Source:MGC

SGC Clone Accession:

Tag:N-terminal His tag with TEV cleavage site

Host:E.coli BL21DE3 R3

Construct

Prelude:

Sequence:

mhhhhhhssgvdlgtenlyfqsmQRGYLLTRNPHLNKDLAFTLEERQQLNIHGLLPSPFNSQEIQVLRVVKNFEHLNSDFDRYLLLM
DLQDRNEKLFYRVLTSDIEKFMPIVYTPTVGLACQQYSLVFRKPRGLFITIHDRGHIAVLNAWPEDVIKAIIVTDGERILGLGDLG
CNGMGIPVGKLALYTACGGMNPQECLPVILDVGTENEELLKDPLYIGLRQRRVRGSEYDDFLDEFMEAVSSKYGMNCLIQFEDFANV
NAFRLLNKYRNQYCTFNDDIQGTASVAVAGLLAALRITKNKLSQITILFQGAGEAALGIAHLIVMALEKEGLPKEKAIKKIWLVDK
GLIVKGRASLTQEKEKFAHEHEEMKNLEATVQEIKPTALIGVAAIGGAFSEQILKDMAAFNERPIIFALSNPTSKAECSAEQCYKIT
KGRAIFASGSPFDPVTLPNGQTLYPGQGNNSYVFPGVALGVVACGLRQITDNIFLTAEVIAQQVSDKHLEEGRLYPPLNTIRDVSL
KIAEKIVKDAYQEKTATVYPEPQNKEAFVRSQMYSTDYDQILPDCYSWPPEEVQ

Vector:pNIC-BSA4

Growth

Medium:

Antibiotics:

Procedure:Cells were grown for 14 hrs at 37°C in 10 mL TB medium, supplemented with Kanamycin (50 µg/mL), and were inoculated into 1 l TB medium with the same antibiotic supplement. The culture was grown at 37°C with shaking at 250 rpm. At an OD of 0.5 cells were induced with 1 mM IPTG and the temperature was lowered to 25°C. Growth was continued for an additional 18 hrs until a final OD of 8-10. Cells were collected by centrifugation, and the cell pellet was stored at -20°C until further use.

Purification

Procedure

Column 1: 1 mL HisTrap FF.

Buffers: Affinity binding buffer: 10mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO₄, 0.5mM TCEP; Affinity wash buffer: 50mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO₄, 0.5mM TCEP; Affinity Elution Buffer: 250mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO₄, 0.5mM TCEP.

Procedure: The supernatant from the last extraction step was loaded on a Akta Express system, consisting of IMAC and gel filtration as consecutive chromatography steps. Before sample loading, columns were equilibrated with the buffers indicated (HisTrap with Affinity binding buffer; gel filtration with Gel Filtration Buffer). Fractions were collected and analysed by mass spectrometry and SDS / PAGE . Peaks after gel filtration were collected, pooled and concentrated using Centricon membrane (30 kDa MW cutoff) c oncentrators.

Column 2 : Superdex S75

Buffers : Gel Filtration Buffer: 10mM pH7.4 Hepes, 500mM NaCl, 5% glycerol, 0.5mM TCEP.

Extraction

Procedure

Pellets were resuspended in approximately 3x volume of lysis buffer, and thawed at 37°C in a water bath. Cells were mechanically disrupted in a French Press, and further sonicated in 10 sec intervals for 2 mins. PEI was added to a final concentration of 0.15%, and DNA was precipitated on ice for 30 mins. The suspension was centrifuged for 17.000 rpm for 30 mins at 4°C, and the supernatant was filtered through a 0.2 um Serum Scrodisc filter. Lysis buffer: 10mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO₄ , 0.5mM TCEP, 1x complete PI EDTA free tablet/50mL.

Concentration:

Ligand

MassSpec:

Crystallization:Column 1: 1 mL HisTrap FF.

Buffers: Affinity binding buffer: 10mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO₄, 0.5mM TCEP; Affinity wash buffer: 50mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO₄, 0.5mM TCEP; Affinity Elution Buffer: 250mM Imidazole, 300mM NaCl, 50mM pH8.0 KHPO₄, 0.5mM TCEP.

Procedure: The supernatant from the last extraction step was loaded on a Akta Express system, consisting of IMAC and gel filtration as consecutive chromatography steps. Before sample loading, columns were equilibrated with the buffers indicated (HisTrap with Affinity binding buffer; gel filtration with Gel Filtration Buffer). Fractions were collected and analysed by mass spectrometry and SDS / PAGE . Peaks after gel filtration were collected, pooled and concentrated using Centricon membrane (30 kDa MW cutoff) c oncentrators.

Column 2 : Superdex S75

Buffers : Gel Filtration Buffer: 10mM pH7.4 Hepes, 500mM NaCl, 5% glycerol, 0.5mM TCEP.

NMR Spectroscopy:

Data Collection:

Data Processing: