

# NQO2

**PDB:**1ZX1

## Revision

**Revision Type:**created

**Revised by:**created

**Revision Date:**created

**Entry Clone Accession:**NQO2A-p002

**Entry Clone Source:**MGC

**SGC Clone Accession:**

**Tag:**N-terminal: His-tag with integrated TEV protease site: mgsshhhhhhssgrenlyfq\*gh(m)

**Host:**BL21(DE3)

## Construct

**Prelude:**

**Sequence:**

mgsshhhhhhssgrenlyfqghMAGKKVLIVYAHQEPKSFNGSLKNNAVDELSRQGCTVTSDLYAMNFEPRATDKDITGTLNPEV  
FNYGVETHEAYKQRSLASDITDEQKKVREADLVIFQFPLYWFSVPAILKGWMDRVLQGFAFDIPGFYDSGLLQGKLALLSVTTGGT  
AEMYTKTGVNGDSRYFLWPLQHGTLHFCGFKVLAQFISFAPEIASEEERKGMAAWSQLQTIWKEEPCTAHWHFGQ

**Vector:**p11

## Growth

**Medium:**

**Antibiotics:**

**Procedure:**The cells were grown at 37°C in 1 L Terrific broth (TB) containing 50 µg/mL ampicillin until reaching OD595 of 0.6. Then the temperature was shifted to 30°C and 0.3 mM IPTG was added to induce the expression. Incubation continued at 30°C for 15 hours before the cells were harvested by centrifugation.

## Purification

**Procedure**

Column 1 : Ni-NTA batch column (Qiagen), 5 mL of 50% slurry in 1.5 x 10 cm column.

Buffers: Washing buffer: 500 mM NaCl, 5% Glycerol, 50 mM Tris-HCl pH 7.5, 30 mM Imidazole, 2 mM TCEP; Elution buffer: 500 mM NaCl, 5% Glycerol, 50 mM HEPES pH 7.5, 250 mM Imidazole, 2mM TCEP.

Procedure: The supernatant was loaded to the Ni-NTA column and equilibrated with the extraction buffer. The column was then washed with 100 mL washing buffer; NQO2 was eluted in 10 mL elution buffer.

Column 2 : Hi Load 16/60 Superdex 200

GF buffer: 100 mM NaCl, 5% Glycerol, 10 mM HEPES pH 7.5, 2mM TCEP

Procedure: Eluted fraction was loaded to the GF column to change the buffer and to achieve homogenous NQO2 for crystallization. Protein was concentrated using a 10000 MW cutoff Amicon Ultra concentration device.

## **Extraction**

### **Procedure**

Extraction buffer: 500 mM NaCl, 5% Glycerol, 50 mM HEPES pH 7.5, 5 mM Imidazole, 2 mM TCEP. Cell pellets from 1 liter were resuspended in 50 mL extraction buffer and then lysed by French Press. The lysate was centrifuged at 18,000 RPM for 1 hour. The cleared lysate was loaded to the Ni-NTA column.

### **Concentration:**

### **Ligand**

### **MassSpec:**

**Crystallization:** Purified NQO2 was concentrated to 12.6 mg/mL and distributed into 50  $\mu$ L aliquots before being flash frozen at -80°C. Crystals were grown using the hanging-drop vapour diffusion technique with 2  $\mu$ L drops. Before set up of the plate, NQO2A was mixed with 50 mM CB1954, 20  $\mu$ M FAD, and then mixed in a 1:1 ratio with reservoir solution (100mM Na-HEPES pH7.0, 2.0 M ammonium sulphate, 5 mM DTT).

### **NMR Spectroscopy:**

### **Data Collection:**

### **Data Processing:**